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Physics/Aerosol Physics

# Fragmentation patterns of particulate organic nitrates in an Aerosol Mass Spectrometer

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<p>Atmospheric aerosols affect the Earth's radiative balance, visibility and human health. Therefore the formation processes and growth of these particles are important and should be studied to understand how human and natural processes affects these processes. One poorly understood and relatively little studied part of aerosols is particulate organic nitrates (pONs). These pONs are mostly formed during nighttime when NO<sub>x</sub>, mainly emitted from fossil fuel combustion and industrial processes, and volatile organic compounds (VOCs), from both natural and anthropogenic sources, reacts in the atmosphere. The quantification of these pONs is still hard due to instrumental restrictions, although much improvement has happened during recent years. One main reason for these challenges is the difficulty to separate inorganic nitrates from organic nitrates with real-time instruments.</p> <p>During this work, we generated pure pON in well controlled laboratory conditions and sampled it with an Aerosol Mass Spectrometer (AMS), an instrument widely used for measuring the chemical composition of atmospheric aerosols. We used four different pON precursors to generate pON. I investigated the fragmentation patterns of pON detected by the AMS, utilizing the high resolution of the newest model of the AMS. As older versions of the AMS has difficulties to separate nitrate-containing organic fragments due to lower resolution than the AMS I used, I was able to study pON mass spectrum with better resolution than anyone before me. I found mass spectral differences for the different pON precursors, and was able to find unique fragments for some of the pON precursors that possibly can be used as marker fragments.</p>			
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# 1. Introduction

Atmospheric aerosols are small solid or liquid particles suspended in air affecting Earth's radiative balance [Russell et al., 1999] and visibility [Charlson, 1969] [Lee et al., 2017]. In addition, they affect negatively on human health [Pöschl, 2005]. Aerosols are emitted either directly to the atmosphere, for example as anthropogenic soot particles from fossil fuel combustion or naturally from oceans as sea salt particles and pollen from vegetation, or are formed secondarily in the atmosphere from condensing vapours. The size, shape, chemical composition, electrical charge and other physical and chemical properties affect how the aerosol particle will interact with its surrounding.

Aerosol particles, ranging in size from a few nm to  $\sim 100\text{ }\mu\text{m}$ , are often measured in number and mass concentration. Even in clean and remote locations (Pallas, Finland), the atmosphere can contain up to 1000 particles per cubic centimeter ( $\text{cm}^{-3}$ ) [Laakso et al., 2003] and in highly polluted urban environments (e.g. New Delhi, India) it can be as high as  $200000\text{ cm}^{-3}$  [Mönkkönen et al., 2004]. The mass concentration has also a wide range, and in the New Delhi study it was as high as  $671\text{ }\mu\text{g m}^{-3}$ . Extremely low particle number concentrations, lower than  $1\text{ cm}^{-3}$ , has been measured in operating rooms in hospitals [Li and Hou, 2003].

In both low and high number concentrations, aerosol particles effect Earth's radiative balance either directly or indirectly. The direct effect is absorption and reflection of solar radiation; dark soot and ash particles absorbs more radiation and warms up the atmosphere while lighter particles scatter solar radiation back to space. Indirect effects come from clouds as all cloud droplets are formed on an aerosol particle, and the aerosol particles' properties will affect the cloud droplets formation and lifetime [Lohmann and Feichter, 2005]. Further, these cloud droplets scatter and absorb solar radiation, affecting the climate. In addition, scattering from aerosol particles affect visibility; higher particle concentration leads to worse visibility. High number and mass concentration, also depending on their chemical composition, is associated with several health issues, including lung and cardiovascular diseases and increased mortality [Brook, 2008] [Abbey et al., 1999] [Lelieveld et al., 2015].

As earlier mentioned, aerosols affect the atmosphere and humans in sev-

eral ways and therefore it is important to understand how different conditions affect aerosol formation. Ever since the industrial revolution in the late 18th century, anthropogenic emissions, especially from fossil fuel combustion, have changed, and still changes, the atmospheres composition [Horowitz and Jacob, 1999][Wuebbles and Jain, 2001][Dignon, 1992] and therefore affect the aerosol formation processes. Several aerosol formation processes are known and well understood, especially the inorganic aerosol formation. The processes to produce organic aerosols are more complex than the inorganics, and not all the processes and their contribution to the total aerosol formation are known.

An example of an aerosol formation process where natural and anthropogenic emissions interacts and produces aerosols is the formation of particulate organic nitrates (pON). pON can form from biogenic (or anthropogenic) emissions and  $\text{NO}_x$  ( $\text{NO} + \text{NO}_2$ ), mostly emitted from fossil fuel combustion and industrial processes. As  $\text{NO}_x$  also is an air pollutant, leading to cardiovascular and respiratory diseases [Chaloulakou et al., 2008], the atmospheric  $\text{NO}_x$  cycle is important to understand. Further, the formation of pON is a sink for  $\text{NO}_x$ , and understanding the whole  $\text{NO}_x$  and pON cycle is essential for air quality control.

The focus in this work is the detection of pON and its separation from inorganic nitrate aerosols via mass spectrometry. This is achieved by investigating the fragmentation patterns of pON from several pON precursors in an Aerosol Mass Spectrometer (AMS), an instrument widely used for measuring the chemical composition of atmospheric aerosols. Unlike previous studies where an AMS has been used, I utilized the newest model of the AMS with its higher resolution to investigate pON-fragments in more detail than earlier possible.



## 2. Theory

### 2.1 Atmospheric aerosols

Atmospheric aerosols are either emitted directly through different sources or secondarily formed in the atmosphere. Sea salt, dust, pollen and ash are examples of primary aerosols while secondary aerosols are formed in the atmosphere via gas to particle conversion. Secondary aerosols can form via inorganic or organic precursors, or a combination of them. The formation of particle phase ammonium ( $\text{NH}_4^+$ ), sulfate ( $\text{SO}_4^{2-}$ ) and nitrate ( $\text{NO}_3^-$ ) from the inorganic gases ammonia ( $\text{NH}_3$ ), sulfur dioxide ( $\text{SO}_2$ ), nitrogen dioxide ( $\text{NO}_2$ ) is fairly well understood. However, the aerosol formation from organic compounds introduces a large uncertainty in the knowledge of their formation mechanisms, properties and composition. As there are estimate up to 100000 measured organic compounds in the atmosphere [Goldstein and Galbally, 2007], the complexity of organic aerosols is enormous.

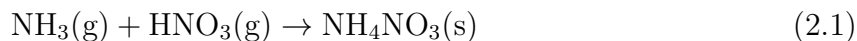
In this work, the emphasis is on the detection of particulate organic nitrates (pON) and its separation from inorganic nitrates via mass spectrometry. Therefore, the following subsections emphasize the formation of both inorganic and organic nitrates, including the  $\text{NO}_x$  cycle and volatile organic compounds (VOCs), without going into detail on other aerosol formation processes.

#### 2.1.1 Inorganic nitrate aerosol

The most commonly measured inorganic aerosol species are ammonium sulfate and ammonium nitrate. Here, we present the formation of ammonium nitrate and other, less common, nitrate salts.

##### 2.1.1.1 Ammonium nitrate

Atmospheric ammonium nitrate ( $\text{NH}_4\text{NO}_3$ , AN) is a product of ammonia ( $\text{NH}_3$ ) and nitric acid ( $\text{HNO}_3$ ):



The main sources of ammonia are anthropogenic emissions [Bouwman et al., 1997]; mainly from agricultural activities, for example fertilizers and domestic animals, although in urban areas, traffic emissions can also be a significant source of ammonia [Phan et al., 2013]. The ammonia emissions have been increasing, and are predicted to increase in the future [Bouwman et al., 1997] [Warner et al., 2017]. Nitric acid is primarily formed, during daytime, when  $\text{NO}_2$  reacts with the OH radical:



### 2.1.1.2 Other inorganic nitrate salts

Other, less common, inorganic nitrate salts, like  $\text{NaNO}_3$  and  $\text{Ca}(\text{NO}_3)_2$  are detected at coastal sites [Allan et al., 2004]. At these sites, sea salt components like sodium chloride ( $\text{NaCl}$ ) and calcium chloride ( $\text{CaCl}_2$ ) can react with nitric acid and form these inorganic nitrate salts.

## 2.1.2 Organic aerosol

Primary organic aerosols (POAs) are emitted both from natural and anthropogenic sources, for example from vegetation as pollen or from vehicles during fossil fuel combustion. Secondary organic aerosol (SOA) is mainly formed via gas to particle conversion from condensable vapours; volatile organic compounds (VOCs) are oxidized and getting low enough vapour pressure to be able to condense and be part of SOA formation and growth. The vast amount of SOA precursors and formation pathways leads to complex chemical composition of SOA. Therefore, it is really hard to chemically identify or separate all the compounds, even with modern measurement techniques. Additional difficulty to detection is introduced by particle phase reactions.

Several oxidation processes for VOCs are known, but in this work we only discuss the organic nitrate formation via VOC oxidation by the nitrate radical. Thereby, the next sections discuss in more detail VOCs and the formation of the nitrate radical.

### 2.1.2.1 Volatile Organic Compounds (VOCs)

A VOC is an organic gas phase compound that has an effective saturation concentration,  $C^*$  (describing the volatility of an compound), larger than  $3 \cdot 10^6 \text{ } \mu\text{g m}^{-3}$

[Donahue et al., 2012]. Carbon monoxide (CO) and carbon dioxide (CO<sub>2</sub>) are not VOCs. If methane is also excluded, 90% of the VOCs are emitted from biogenic and 10% from anthropogenic sources [Seinfeld and Pandis, 2016]. Globally, the largest non-methane biogenic VOC (BVOC), emitted from the vegetation, measured in mass concentration, is isoprene (C<sub>5</sub>H<sub>8</sub>) (~ 70%), followed by monoterpenes (C<sub>10</sub>H<sub>16</sub>) (~ 11%) [Sindelarova et al., 2014]. To monoterpenes belongs for example  $\alpha$ -pinene,  $\beta$ -pinene and limonene, and they are the major BVOCs in the boreal forest in the northern hemisphere [Rinne et al., 2009]. Anthropogenic VOCs are mainly alkanes, aromatics and alcohols emitted for example from combustion processes and solvents [Theloke and Friedrich, 2007].

When emitted to the atmosphere, VOC are primarily oxidized by ozone (O<sub>3</sub>), the hydroxyl radical (OH) and nitrate radical (NO<sub>3</sub>). VOCs can be oxidized several times, lowering their volatility until they condense and form SOA. During daytime, OH radical is the dominant oxidant, while during night-time the NO<sub>3</sub> radical plays a major role.

The VOCs used in this work are presented in more detail later on in Section 4.1.2.

#### 2.1.2.2 NO<sub>x</sub> and NO<sub>3</sub> chemistry

The main sources of tropospheric NO<sub>x</sub> (NO<sub>x</sub> = NO + NO<sub>2</sub>) are fossil fuel combustion and industrial processes [Seinfeld and Pandis, 2016], contributing nearly 60% of the total NO<sub>x</sub> emissions. Natural sources, like soils and lightnings, accounts for 23% of the total NO<sub>x</sub> emissions. The NO<sub>x</sub> chemistry differs a lot during day- and nighttime. During daytime, NO is rapidly oxidized by ozone (O<sub>3</sub>) to NO<sub>2</sub> and NO<sub>2</sub> is photolysed back to NO. During nighttime, when no sunlight is present, NO is converted to NO<sub>2</sub> by:



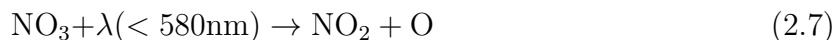
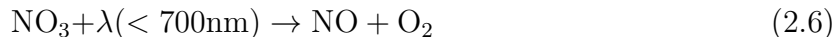
Further, the nitrate radical (NO<sub>3</sub>) is produced from NO<sub>2</sub> oxidation by O<sub>3</sub>:



Reaction 2.4 is the only direct atmospheric source of NO<sub>3</sub>, an indirect source is through thermal decomposition of N<sub>2</sub>O<sub>5</sub>:



Reaction 2.5 reaches an equilibrium within a few minutes [Seinfeld and Pandis, 2016] during nighttime. During daytime, any nitrate radicals would decompose either via photolysis or via reaction with NO within seconds:



Hence, the nitrate radical is important mainly during nighttime chemistry as it is a strong oxidant for a wide range of VOCs. Especially fast reactions with NO<sub>3</sub> occurs with unsaturated compounds (containing at least one carbon-carbon double or triple bond) [Atkinson and Arey, 2003], for instance alkenes. The low volatility compounds formed during oxidation of BVOCs by the NO<sub>3</sub> radical, producing SOA, is a direct link between biogenic carbon in atmospheric aerosols and anthropogenic pollution (NO<sub>x</sub>).

### 2.1.2.3 Particulate organic nitrate (pON)

Although it is known that organic aerosols are a major part of atmospheric aerosols, there are still large uncertainties related to the sources of these organic compounds [Hallquist et al., 2009]. Modelling studies [Spracklen et al., 2011] show that reactions between anthropogenic emissions and BVOCs could play an important role in the global SOA budget. One candidate for the source of this organic mass is particulate organic nitrate (pON). For example, [Kiendler-Scharr et al., 2016] presented results from measurements around Europe, during several seasons, that pON could account for up to 44% of the submicron nitrate aerosol. In addition, [Lee et al., 2016] estimated that pON could account for 8% of the total nighttime OA mass in the south-east United States.

Organic nitrates, in the gas phase, are formed by oxidation of VOCs by NO<sub>3</sub> radicals, adding a nitrate (-ONO<sub>2</sub>) functional group to the VOC and lowering their vapour pressure. Further, pON is formed when organic nitrates in the gas phase are partitioning to the particulate phase and form SOA.

## 2.2 Challenges in aerosol measurements

Besides the vast chemical complexity for organic aerosols, another major reason for difficulties in chemical characterization of aerosols is the wide range of sizes. A particle with a diameter of 1 nm and 1 μm will have a mass of approximately 10<sup>-21</sup> g (zeptograms) and approximately 10<sup>-12</sup> g (picograms), respectively. In atmospheric studies,

the time-dependent evolution of the aerosol particles is often an aspect of interest, favoring real-time instruments that do not separately collect, store and analyze the samples.

For particulate organic nitrates (pON) there is yet no accurate real-time analysis method to detect pON and quantify them. Off-line methods can give more specific information of the chemical composition and structure, while real-time instruments usually gain the time-resolution by giving up the chemical specificity.

In this work, we used an Aerosol Mass Spectrometer (AMS) designed and developed by Aerodyne Research, Inc. (ARI). The AMS is a real-time instrument that can measure size-resolved chemical composition of non-refractory submicron particles from ambient pressure (for more detailed description, see Section 3.1). The AMS uses thermal vaporization and electron impact ionization (70 eV) to maximize the range of detected aerosol species, but loses, via fragmentation of the original molecules, a lot of chemical information due to this hard ionization method. Most of the organic and inorganic nitrates fragment into  $\text{NO}^+$  and  $\text{NO}_2^+$  within the AMS. Furthermore, the ratio of  $\text{NO}^+$  and  $\text{NO}_2^+$  ( $\text{NO}_x^+$  ratio) differs depending if the nitrate comes from organic or inorganic compounds, and this could potentially be used to quantify pON from ambient data [Farmer et al., 2010] by

$$x = \frac{(R_{\text{obs}} - R_{\text{AN}})(1 + R_{\text{pON}})}{(R_{\text{pON}} - R_{\text{AN}})(1 + R_{\text{obs}})} \quad (2.9)$$

Where  $x$  is the fraction of the total nitrate signal due to organic nitrates.  $R$  is the  $\text{NO}_x^+$  ratio for ammonium nitrate (AN), particulate organic nitrate (pON) and the ambient observed ratio (obs).

The main source of inorganic nitrate in the atmosphere is ammonium nitrate, except at coastal sites where  $\text{NaNO}_3$  and  $\text{Ca}(\text{NO}_3)_2$  can contribute significantly to the inorganic nitrate portion, increasing the  $\text{NO}_x^+$ -ratio [Allan et al., 2004]. Several studies has shown that the  $\text{NO}_x^+$  ratio for pure AN is ranging from 1 to 3 while it is way higher for pON, ranging often from 5 to 10 (see Figure 5.10 for ratios from previous studies).

There are still several problems for using the  $\text{NO}_x^+$  ratio for pON quantification. First, the  $\text{NO}_x^+$  ratio for pON differs a lot depending on the precursors available on different sites and different precursors give different  $\text{NO}_x^+$  ratios. Additional uncertainty is also introduced by an organic fragment,  $\text{CH}_2\text{O}^+$ , which is often present in ambient measurements at the same unit mass as the  $\text{NO}^+$  fragment. If the resolution of the instrument is not good enough to separate these two fragments, underestimation of the organic fragment can lead to significant overestimation of pON. If the pON concentrations are too low, this approach is not capable of quantifying pON, restricting this method to locations with large fraction of pON. Additionally, this method does

not work at costal sites where  $\text{NaNO}_3$  or  $\text{Ca}(\text{NO}_3)_2$  are present.

## 3. Instruments

In this section, the working principles of the three main instruments (AMS, PAM, AAC) for this work is presented in more detail, while other instruments are described shortly in the methods section when mentioned.

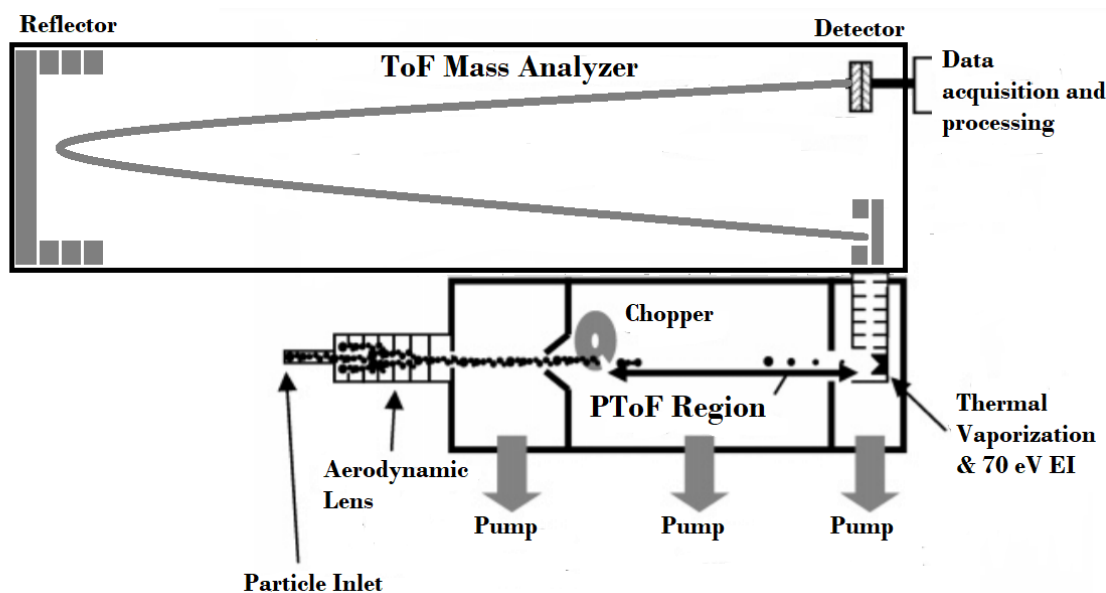
### 3.1 Aerosol Mass Spectrometer (AMS)

The Time of Flight Aerosol Mass Spectrometer (ToF-AMS) is a real-time instrument that can measure size-resolved chemical composition of submicron particles from ambient pressure. The working principles of a Long-ToF-AMS (L-ToF-AMS), the main instrument in this work, is similar to the High Resolution (HR) ToF-AMS (DeCarlo et al. 2006), but it is mounted with a longer ToF-chamber enabling over two times higher resolution. There are different versions of the AMS (described in Section 3.1.5); all of them are based almost on the same techniques, but some differences related to technique development and achieving desired functionalities are found. The four main parts of the AMS are 1) the aerosol inlet, where the sampling takes place, 2) the particle sizing chamber, where the size of the particles are measured, 3) the vaporisation and ionisation chamber, where the solid or liquid particles are vaporised and ionised and 4) mass analyser, where the chemical composition of the particles are measured via mass spectrometry (quadrupole or time-of-flight mass analysers). The AMS is developed and manufactured by Aerodyne Research Inc., Billerica, Massachusetts, United States.

The schematics of the L-ToF-AMS is shown in Figure 3.1

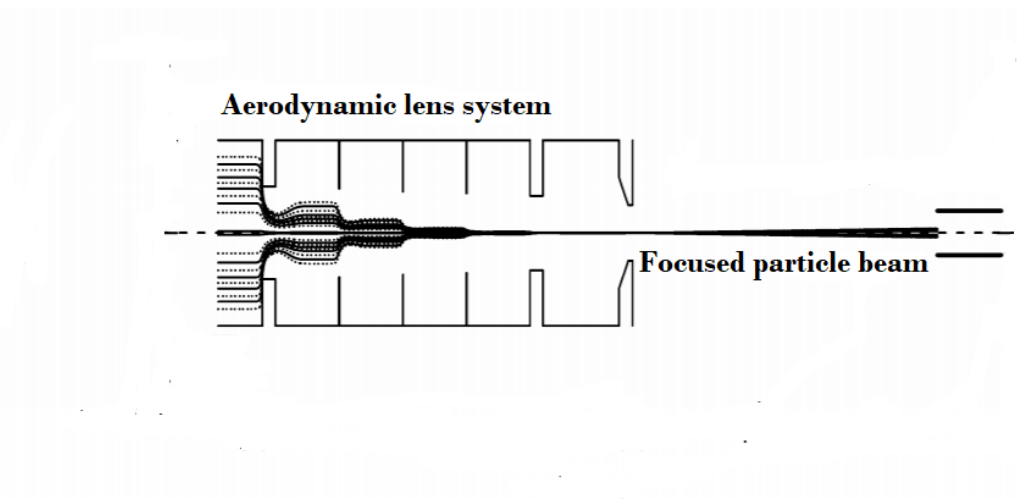
#### 3.1.1 Inlet and aerodynamic lens

The AMS samples air through a critical orifice (diameter = 100  $\mu\text{m}$ ), which drops the pressure from ambient (101.325 kPa) to under 300 Pa [Canagaratna et al., 2007]. The flow is now led through an aerodynamic lens system, which focuses the particles to a narrow beam with a diameter of approximately 1 mm. The transmission efficiency of the aerodynamic lens is a function of particle diameter, and has been studied both experimentally and theoretically for example by [Liu et al., 2007]. Depending on the



**Figure 3.1:** The schematics of the L-ToF-AMS (Figure from [DeCarlo et al., 2006], modified from the HR-ToF-AMS schematic)

definition of transmission, the AMS is reported to detect particles with an aerodynamic diameter between 40 nm and 1  $\mu\text{m}$ . Particle trajectories inside the lens system is shown in Figure 3.2.



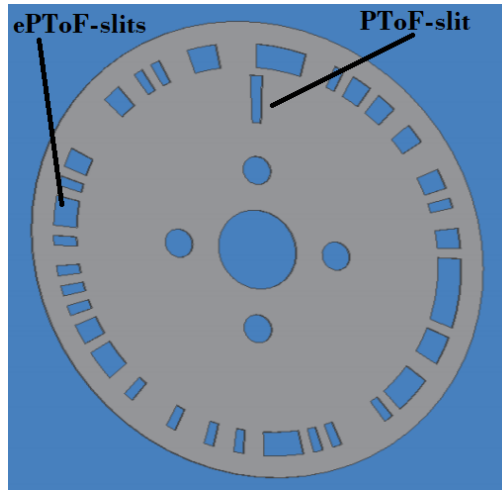
**Figure 3.2:** Particle trajectories inside the aerodynamic lens, focusing the particles to a narrow beam (figure from Zhang et al. 2004, modified)

### 3.1.2 Particle sizing chamber

When exiting the aerodynamic lens, and entering the sizing region of the instrument, the flow undergoes a supersonic expansion where the particles are accelerating and



achieving a velocity-distribution depending on their aerodynamic size. The particle size is calculated from the particle time of flight (PToF) across a well-defined distance ( $\sim 30$  cm) between a chopper and the ionization chamber (PToF region). The beginning of the flight time starts at a mechanical chopper, mounted after the aerodynamic lens and consisting of 32 slits on the chopper wheel ( $\sim 5$  cm in diameter, see Figure 3.3), chopping the particle beam with a 90 Hz frequency. The particle flight time ends at the mass spectrometric detection providing chemically resolved particle size-data, although the PToF region ends before it. This is possible because the particle flight time is on the millisecond scale ( $\sim 3$  ms) and the vaporization, ionization and MS-detection is on the microsecond scale ( $\sim 50$   $\mu$ s). The original PToF chopper had only 1 slit (instead of 32 slits) chopping the particle beam, but newer models of the AMS uses this chopper with 32 slits and is called efficient particle time of flight (ePToF). The ePToF, utilizing a Hadamard transform [Zare et al., 2003], reaches higher transmission ( $\sim 50\%$ ) of particles compared to the old PToF version that had only  $\sim 2\%$  transmission. The AMS used in this work was equipped with the ePToF chopper.



**Figure 3.3:** Chopper from AMS particle sizing chamber. The one inner slit (PToF-slit) is used for PToF-mode and all of the outer slits (ePToF-slits) are used to chop the particle beam during ePToF-mode. The one big circular hole and the four smaller in the middle are used for mounting the chopper to the axis that moves the chopper between the different modes (Figure from [Aerodyne, ], modified)

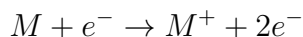
Besides the particle sizing measurements, the chopper has another very important function. It is used to measure the background signal from the air that has not been removed from the chamber by the pumps. The chopper has four different modes that can be used during measurements: open, closed, PToF and ePToF. In open-mode, the chopper is moved away from the particle beam, allowing all particles and air to reach the vaporisation and ionisation chamber. In closed-mode, the chopper is blocking all particles and used to calculate the signal obtained from the ionised background gases.

The difference of open- and closed-mode is used as the signal from the sampled aerosols. PToF and ePToF is used as earlier described. Usually, the AMS is operated by altering the open/closed-modes and ePToF (or PToF) with different frequencies, depending on wanted amount of particle size and chemical composition data.

For this work, no ePToF data were analyzed because we selected well-defined monodisperse aerosols with an AAC (described later on in Section 3.2) before sampling it with the AMS.

### 3.1.3 Vaporisation and ionisation

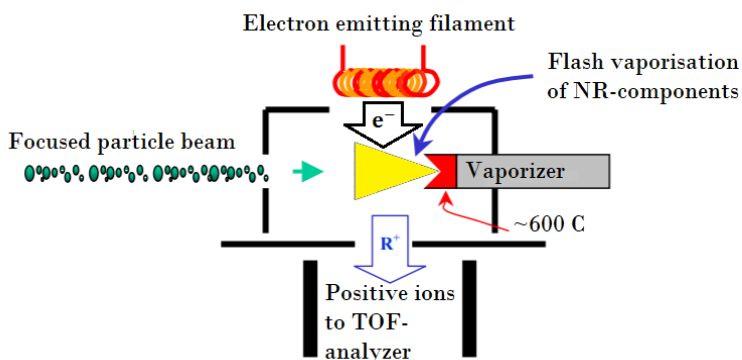
After the PToF region, the particles enters the vaporization and ionization chamber where they are flash vaporised by impact on an  $\sim 600^{\circ}\text{C}$  porous tungsten surface. The vaporizer has a diameter of  $\sim 4$  mm and is formed as an inverted cone to reduce unwanted particle bounce that reduces the collection efficiency of particles [Middlebrook et al., 2012]. Due to the selected vaporisation method only "non-refractory" (NR) species are vaporised (defined as those species that are vaporised in these condition), including organics and most nitrate and sulfate salts. This means that sea salt, metals and black carbon are not detected by the AMS [Canagaratna et al., 2007]. The gaseous molecules are now ionised through electron ionisation (EI, formerly referred as electron impact ionisation). Schematics of the vaporisation and ionisation chamber is shown in Figure 3.4. EI is based on shooting energetic electrons (70 eV in the AMS) on neutral vapors and transferring some of the kinetic energy to the sample molecules when colliding. If the collision is successful, the transferred energy exceeds the ionisation energy of the molecule and the molecule loses an electron and forms a molecular ion (a positive radical ion):



EI is an easy ionisation method with good repeatability and it is non-selective, ionizing both organics and inorganics. As the AMS uses standard 70 eV EI, its mass spectra can be compared to the National Institute of Standards and Technology (NIST) mass spectral database [Linstrom and Mallard, 2001] and an intercomparison of AMS and NIST spectra from several chemical species shows that the mass spectra are very similar except from some small differences for long-chained organics [Alfarra, 2004]. The electron used in EI are generated by thermionic emission from a tungsten wire (filament), which is resistively heated up to  $\sim 2000^{\circ}\text{C}$ . Extracted electrons are then accelerated by a voltage to achieve wanted 70 eV electrons. However, if the accelerated electrons have lower or higher energy than 70 eV, it may result in different fragmentation and loss of resolution and mass accuracy [Gross, 2004]. The downside of EI is the fragmentation

of the parent molecule, losing some chemical information of them. Furthermore, identification of the original molecule structure is not possible due to no prior separation method, only giving the chemical composition of the fragments. In addition, due to the large scale of possible compounds in the aerosol particle, especially organics, the detected mass spectra is a mixture of fragments from all molecules and makes it practically impossible to identify the chemical composition of any specific molecules from ambient aerosols. The fragmentation can, however, be used to identify some species if they have a specific fragmentation pattern. For example, the nitrates in both organic nitrates and inorganic nitrates fragments mostly to  $\text{NO}^+$  and  $\text{NO}_2^+$ , but the ratio of these (the  $\text{NO}^+/\text{NO}_2^+$ -ratio, hereafter  $\text{NO}_x^+$  ratio) is different for organic and inorganic nitrates and can be utilized for quantification of organic nitrates [Farmer et al., 2010]. More of the  $\text{NO}_x^+$  ratio previously Section 2.2.

The detection scheme and calibration method of the AMS allows quantitative detection of ambient aerosol [Jimenez et al., 2003]. The AMS is usually calibrated with  $\text{NH}_4\text{NO}_3$  and  $(\text{NH}_4)_2\text{SO}_4$ , mainly because nitrate and sulfate is common aerosol species and vaporises easily in the vaporiser.



**Figure 3.4:** Vaporisation and ionisation chamber. Particles first hit the 600°C porous tungsten vaporiser and are then ionised by 70 eV electrons emitted from a filament. Positive ions are then directed to the ToF mass analyser (Figure from [Canagaratna et al., 2007], modified)

### 3.1.4 Mass analyser

The mass analyser used in most AMS versions are different length of orthogonal acceleration reflector time of flight-analysers (oaReToF, hereafter simply ToF). The basic principle for a ToF is simple: ions with different mass to charge ratio ( $m/z$ ) and same initial kinetic energy, moves with different velocities in vacuum, and by knowing the length of the flight path, it is possible to calculate the  $m/z$  for the detected ions. We assume a particle at rest with the mass  $m$  and the charge  $q$  (which is equal to an integer

number  $z$  of elemental charges,  $q = ez$ ). Before entering the ToF chamber, the ions are given kinetic energy by moving through a voltage  $U$  and the energy uptake there ( $E_{\text{el}}$ ) is converted to kinetic energy ( $E_{\text{kin}}$ ):

$$E_{\text{el}} = qU = ezU = \frac{1}{2}mv^2 = E_{\text{kin}} \quad (3.1)$$

The ions have now the following velocity:

$$v = \sqrt{\frac{2ezU}{m}} \quad (3.2)$$

and their flight time along a field free distance  $L$  is:

$$t = \frac{L}{v} = \frac{L}{\sqrt{\frac{2ezU}{m}}} \quad (3.3)$$

Each instrument has their well-defined values for  $U$  and  $L$ , and by measuring the flight time  $t$ , the ratio  $m/z$  is:

$$\frac{m}{z} = \frac{2eUt^2}{L^2} \quad (3.4)$$

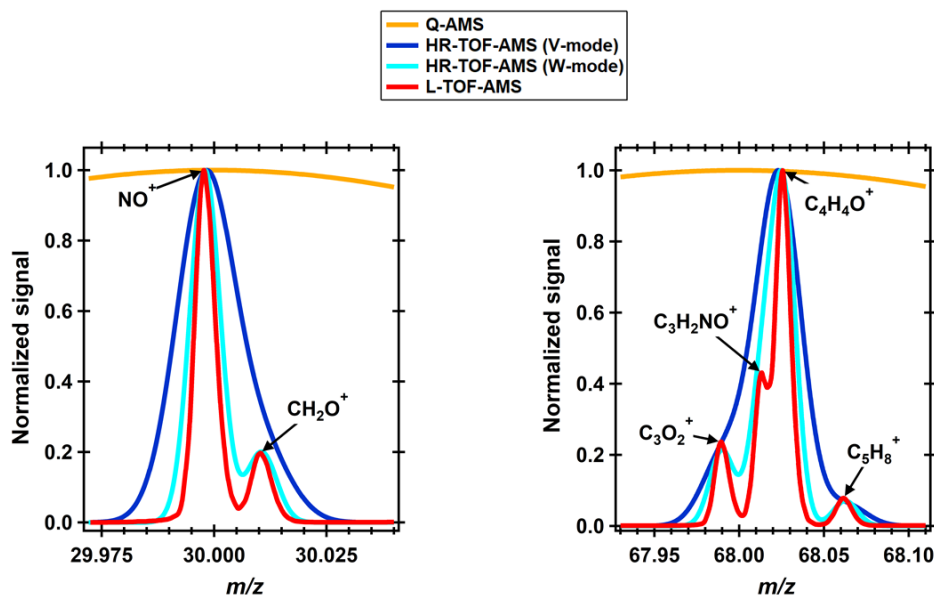
The reflectors working principle is following: behind the field-free region a retarding electric field is located and works as a mirror that changes the ions flight direction and lengthens the flight path. Differences in the initial energy for ions with same  $m/z$ -ratio, resulting in different flight time, is corrected by the reflector and increasing the resolution remarkably. This is achieved by the retarding electric field: ions with higher kinetic energy will penetrate deeper into the electric field than ions with lower energy and thus spend more time in the electric field and thereby correcting the flight time. In all AMS-versions, excluding the HR-ToF-AMS, V-mode stands for normal one-reflector mode but in the HR-ToF-AMS there is also a W-mode. In the W-mode, the ions exiting the reflector are sent back to the reflector one more time by a hard mirror (instead of going to the detector), lengthening the flight path further. The W-mode has higher resolution than V-mode, but the V-mode is more sensitive because more ions are lost in the W-mode due to lateral broadening and fewer ions reaching the detector.

In an orthogonal acceleration ToF, pulses of ions are extracted orthogonally to the ToF from the ion beam. The main advantages of oaToF is high sensitivity, high acquisition rate and compact design [Gross, 2004]. The flight path of the ions inside the ToF is seen in Figure 3.1.

In this work, no data acquisition or processing nor ion detection details are presented but can be found from [DeCarlo et al., 2006] [Sueper and collaborators, ].

### 3.1.5 Different AMS versions

There have been several different versions of the AMS, and the main difference has been the length the ToF chamber. However, the first AMS, the Q-AMS, had a quadrupole instead of ToF chamber as mass analyser and had only unit mass resolution (UMR). Its time resolution were also significantly worse. AMS versions using a ToF, there are different versions (listed with increasing resolution): the compact (C-ToF), HR-ToF and L-ToF AMS. In the HR-ToF-AMS, there are the V- and W-mode for further increase of resolution. In Figure 3.5, the resolution a HR-ToF-AMS (V- and W-mode), a L-ToF-AMS and a Q-AMS is compared. From that figure it is clear that the L-ToF-AMS has the highest resolution and is able of separating fragments significantly better than the HR-ToF-AMS, even in W-mode. The high resolution of the L-ToF-AMS is utilized for identifying nitrogen-containing organic fragments in this work. In addition to the different AMS versions, an Aerosol Chemical Speciation Monitor (ACSM) has been developed by Aerodyne Inc. [Ng et al., 2011]. The ACSM is basically a simpler version of the AMS, but without the particle-sizing part. While the AMS is mainly meant for shorter measurements, the ACSM is developed for long-time (years) monitoring of submicron mass concentration and chemical composition. The original ACSM uses a quadrupole mass analyser, but some newer models are equipped with a ToF analyser.



**Figure 3.5:** Comparison of the resolution of different AMS versions. The L-ToF-AMS data is from an actual experiment done during this work, the rest of the data is produced artificially knowing the resolution of each instrument version.

## 3.2 Aerodynamic Aerosol Classifier (AAC)

The aerodynamic aerosol classifier (AAC) is an instrument for selecting monodisperse aerosols from polydisperse aerosol population and it classifies aerosols according to their aerodynamic diameter [Tavakoli and Olfert, 2013]. The schematics of an AAC is shown in Figure 3.6. It consists of two concentric cylinders, rotating with same rotational speed and direction. When particles enter the AAC, from the aerosol flow ( $Q_a$ ), they are guided through a small gap leading to a particle free sheath flow ( $Q_{sh}$ ) between the two concentric cylinders. The particles experience a centrifugal and drag force in the radial direction due to the rotating cylinders, but are also carried in the axial direction due to the sheath and aerosol flow. Particles with a specific aerodynamic diameter are now exiting the AAC with the sample flow ( $Q_s$ ). Particles with larger aerodynamic diameter hit the inner wall before the exit and particles with smaller aerodynamic diameter hit the inner wall after the slit or exit the AAC with the exhaust flow ( $Q_{exh}$ ). Particle trajectory and the flows inside the AAC is presented in Figure 3.6. The AAC is manufactured by Cambustion Ltd., Cambridge, United Kingdom.

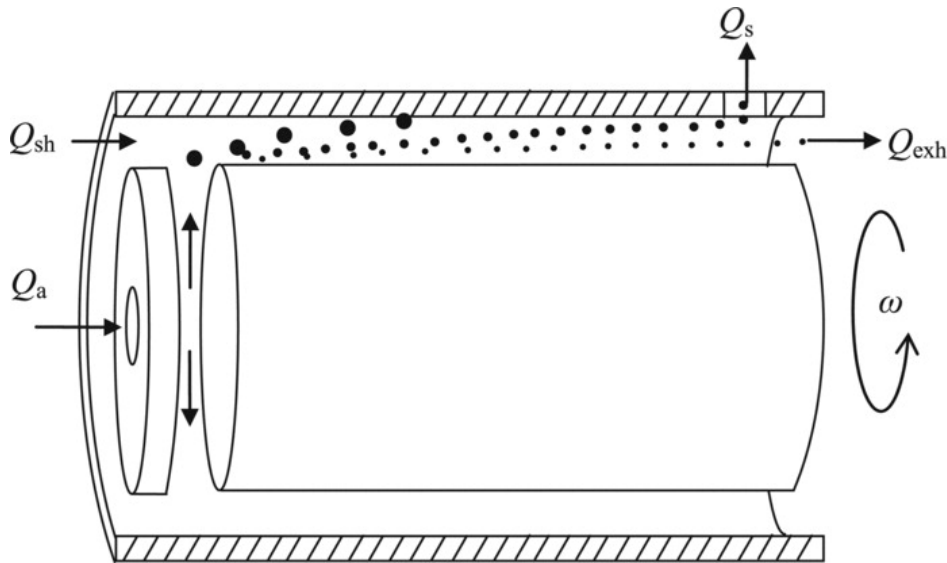


Figure 3.6: Schematics of the AAC [Tavakoli and Olfert, 2013])

## 3.3 Potential Aerosol Mass Oxidation Flow Reactor (PAM-OFR)

[Kang et al., 2007] defined a new concept of Potential Aerosol Mass (PAM) as the maximum aerosol mass that the oxidation of precursor gases can produce. They also

developed a small flow reactor to investigate the PAM concept. The PAM oxidation flow reactor (PAM-OFR, hereafter PAM) is a simple, small, flow-through chamber where atmospheric oxidation processes can be studied in laboratory or in field. The PAM provides a highly oxidizing environment, simulating atmospheric oxidation processes with typical atmospheric time-scales from hours up to several days in only a few minutes. The PAM is capable of shortening the typical long atmospheric oxidation-processes to only a few minutes due to extremely high amounts of oxidants introduced to the system. The PAM chamber itself is a small (13 L) cylindrical chamber, where huge amounts of oxidants (e.g  $\text{O}_3$ ,  $\text{OH}$  or  $\text{NO}_3$ ) and a precursor gas is introduced and then rapidly oxidized. These oxidized compounds can then form SOA that can further be investigated, for example with an AMS. Some version of the PAM has UV lamps to simulate daytime atmospheric conditions, but for this work we used a dark PAM chamber to simulate nighttime conditions and promote  $\text{NO}_3$  radical chemistry.





## 4. Measurements and data analysis

### 4.1 Measurements

All of the measurements in this work was done during an ACSM-intercomparison at the Aerosol Chemical Monitor Calibration Centre (ACMCC) in Paris, France, during November and December 2018. During the measurements, pON was generated in a PAM from the reaction of  $\text{NO}_3$  radicals and a VOC. We used four different pON precursor VOCs (described in Section 4.1.2) and investigated the detection and fragmentation of the generated pON in the L-ToF-AMS and ACSMs. One of the main goals during the ACSM-intercomparison was to utilize the high resolution of the L-ToF-AMS to evaluate the quantification of pON with the low-resolution ACSMs.

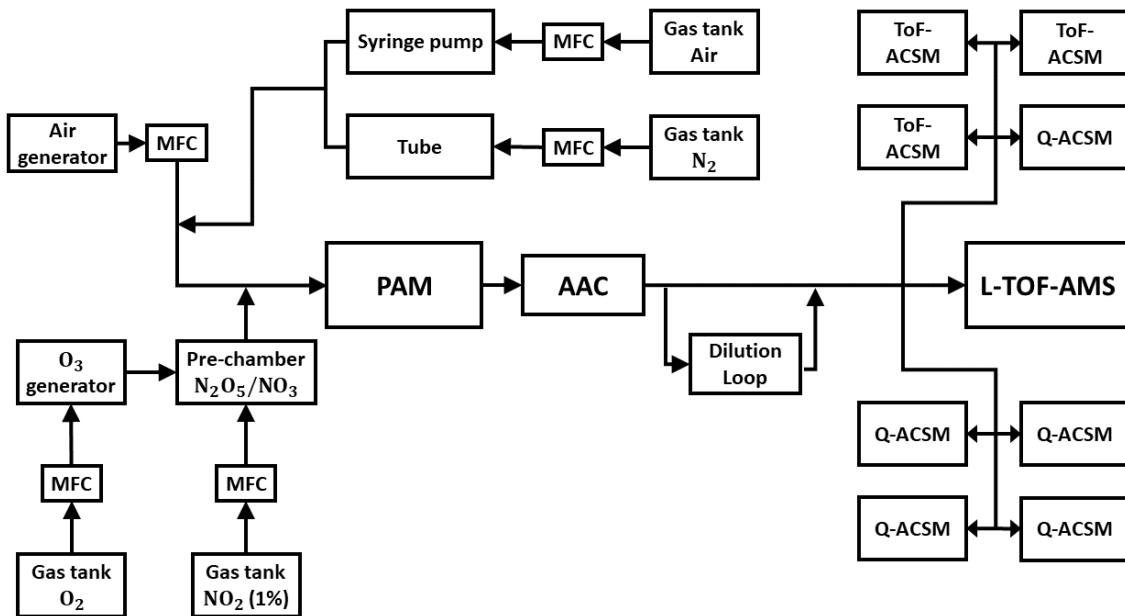
#### 4.1.1 Experimental setup

The experimental setup used during the pON experiments is presented in Figure 4.1. The experimental setup was designed and set up by the ACMCC crew. The pON precursors were either introduced via a syringe pump (liquids) or a tube (powder) together with air or  $\text{N}_2$  gas, flow rates in the lines were controlled by mass flow controllers (MFCs). The generation of nitrate radicals was done by  $\text{O}_3$  and  $\text{NO}_2$  reactions (Reaction 2.4 and 2.5).

Produced pON exiting the PAM was then size-selected with the AAC and monodisperse aerosols were then sampled with the L-ToF-AMS and the 8 ACSMs. The maintenance and operating of the L-ToF-AMS during the experiments was my responsibility.

#### 4.1.2 pON precursors

We used four pON precursors (VOCs) during this experiment, two biogenic (limonene and  $\beta$ -pinene) and two anthropogenic (guaiacol and acenaphthylene). The chemical structure of all compounds are shown in Figure 4.2. Only acenaphthylene was introduced as powder in the system, rest of the compounds were liquids. The SOA formation



**Figure 4.1:** Experimental setup. pON precursors and  $\text{NO}_3$  radicals are introduced to the PAM chamber. Before sampling the aerosols with the L-ToF-AMS and the ACSMs, monodisperse aerosol population is generated with the AAC.

in the PAM was driven by  $\text{NO}_3$  oxidation even though some  $\text{O}_3$  did enter the PAM, mainly because the  $\text{NO}_3$  radicals reacts faster than  $\text{O}_3$  with the investigated VOCs. The atmospheric lifetimes of all four precursors with respect to the three main atmospheric oxidants ( $\text{NO}_3$ ,  $\text{O}_3$  and  $\text{OH}$ ) are presented in Table 4.1. From there we can easily see that the VOCs used are oxidized way faster by the nitrate radical than by  $\text{O}_3$  in nighttime conditions and as we simulated nighttime conditions we can assume that the SOA formation in the PAM was  $\text{NO}_3$  driven.

#### 4.1.2.1 Guaiacol

Guaiacol (2-methoxyphenol) is a semivolatile gas phase methoxyphenol produced during biomass burning, both natural fires and human related fires such as residential wood combustion [Lauraguais et al., 2016], and can be used as a tracer for atmospheric wood smoke emissions due to its unique production from biomass burning [Hawthorne et al., 1988] [Simoneit et al., 1993].

In an oxidation flow reactor (OFR) experiment, [Liu et al., 2019] produced organic nitrates by oxidizing guaiacol with  $\text{OH}$ -radicals in presence of  $\text{NO}_2$  and reported a  $\text{NO}_x^+$  ratio of 4.02-6.25 detected with a HR-ToF-AMS.

**Table 4.1:** Atmospheric lifetimes of VOCs used in the experiments with respect to reactions with  $\text{NO}_3$  (during nighttime),  $\text{O}_3$  (24h average) and  $\text{OH}$  (during daytime).

Oxidant	$\text{NO}_3$ (nighttime)	$\text{O}_3$ (all day)	$\text{OH}$ (daytime)
Acenaphthylene	7 min <sup>a</sup>	1 h <sup>a</sup>	1.3 h <sup>a</sup>
Guaiacol	74 s <sup>b</sup>	11.8 days <sup>c</sup>	2.3 h <sup>b</sup>
Limonene	9 min <sup>d</sup>	36 min <sup>d</sup>	2 h <sup>d</sup>
$\beta$ -pinene	50 min <sup>d</sup>	18 h <sup>d</sup>	3.6 h <sup>d</sup>

<sup>a</sup> [Zhou and Wenger, 2013]  $[\text{NO}_3] = 24$  ppt,  $[\text{O}_3] = 280$  ppb,  $[\text{OH}] = 2 \cdot 10^6 \text{cm}^{-3}$

<sup>b</sup> [Lauraguais et al., 2016]  $[\text{NO}_3] = 20$  ppt,  $[\text{OH}] = 1.6 \cdot 10^6 \text{cm}^{-3}$

<sup>c</sup> [Zein et al., 2015]  $[\text{O}_3] = 100$  ppb

<sup>d</sup> [Winer et al., 1984]  $[\text{NO}_3] = 10$  ppt,  $[\text{O}_3] = 30$  ppb,  $[\text{OH}] = 1 \cdot 10^6 \text{cm}^{-3}$

#### 4.1.2.2 Acenaphthylene

Acenaphthylene is a polycyclic aromatic hydrocarbon (PAH) mainly emitted from fossil fuel combustion, both from liquid and solid fuels [Ravindra et al., 2008]. Under normal atmospheric conditions acenaphthylene exists in gas phase and it is highly reactive to atmospheric oxidants due to its carbon-carbon double bond.

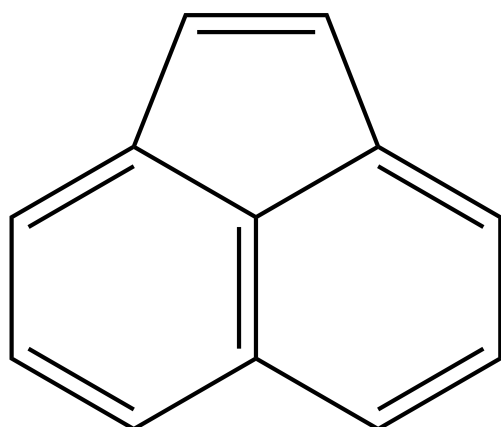
The purity of the acenaphthylene used in this experiment was only 75%, the rest was acenaphthene that differs from acenaphthylene only by one carbon-carbon double bond less (chemical structure shown in Figure 4.2e).

#### 4.1.2.3 Limonene

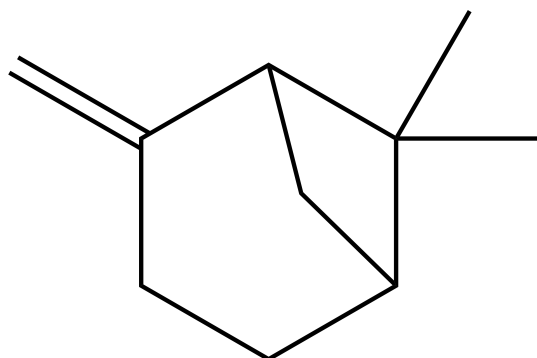
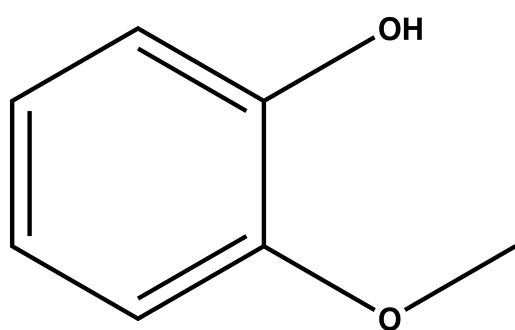
Limonene is one of the most emitted monoterpenes in the nature (Sakulyanontvittaya et al. 2008 and [Guenther et al., 2012], but it is also used in several air fresheners and cleaning products [Wainman et al., 2000] due to its lemon/citrus scent. Limonene possesses two double bonds and oxidizes rapidly in the atmosphere and can easily form products that can further form SOA. Earlier reported  $\text{NO}_x^+$  ratios from limonene +  $\text{NO}_3$  systems are  $\sim 6.67$  [Fry et al., 2011] and 6.3 [Boyd et al., 2017].

#### 4.1.2.4 $\beta$ -pinene

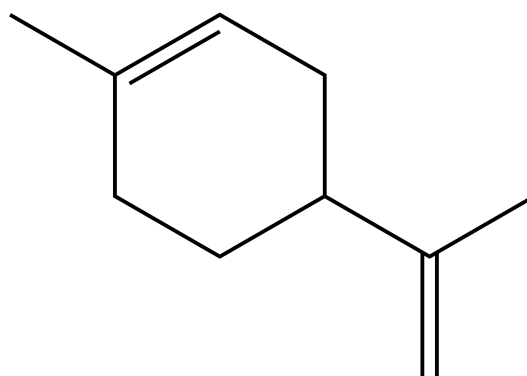
$\beta$ -pinene is a more abundantly emitted monoterpene than limonene [Guenther et al., 2012] and a modelling study from 2005 (Russel and Allen) suggested that up to 20% of nocturnal SOA is formed through  $\beta$ -pinene +  $\text{NO}_3$  reactions. Beta-pinene +  $\text{NO}_3$  systems has been frequently studied and  $\text{NO}_x^+$  ratios from 6.3 to 10 has been reported [Fry et al., 2009], [Nah et al., 2015],[Boyd et al., 2015], [Boyd et al., 2017].



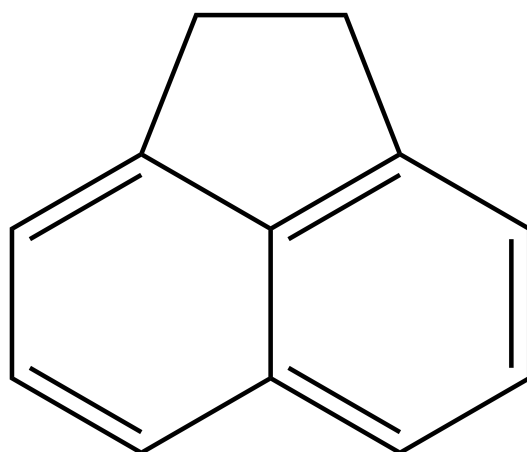
(a) Acenaphthylene

(b)  $\beta$ -pinene

(c) Guaiacol



(d) Limonene



(e) Acenaphthene

**Figure 4.2:** Chemical structures of all four precursors and acenaphthene (impurity for acenaphthylene).

## 4.2 Data analysis

In this work, I will not go into the details of AMS data analysis, as this would also require a more detailed instrumentation description and thereby only the main analysis parts are mentioned here. However, the data analysis is a crucial part for correct identification of the fragments detected with the AMS.

Firstly, a  $m/z$  calibration is performed: the time-of-flight data from the raw spectra is converted to  $m/z$  values. This is done by choosing a few peaks with known  $m/z$  values and using them to calculate the  $m/z$  values for the rest of the data. There are few criteria for the peaks used in the  $m/z$  calibration. One must know exactly which ions are used so that the correct  $m/z$  value are set for the peak. The ions used for the  $m/z$  calibration should also cover a wide range of masses, so that peaks at both low and high masses are pinpointed correctly to the right  $m/z$  values.

After the  $m/z$  calibration a baseline is calculated. This takes into account the background signal that is measured with the chopper (open and closed mode), described previously in Section 3.1.2. Next is the peak shape and width determination. During this step, a function is fitted to the measured signal (see Appendix A for visualisation of this). This step is also crucial for high resolution data analysis: with wrong peak shape and width, small-signal ions close to bigger peaks can be lost during this step. When all of the peak shape parameters are determined, one must still manually go through all ions and choose which should be fitted. After this there are still a few steps before the final processed data is ready.

A more detailed description of high resolution AMS data analysis can be found in [DeCarlo et al., 2006].

All of the data analysis for AMS data is done with two AMS analysis packages implemented in the data analysis software Igor Pro by WaveMetrics Inc. The two AMS packages, developed by Aerodyne, is SQUIRREL (SeQUential Igor data RetRiEvaL) and PIKA (Peak Integration by Key Analysis). The software versions used in this work was Igor Pro 6.37, SQUIRREL 1.62A and PIKA 1.22A.

All of the data analysis for this work was done by me.



# 5. Results and discussion

In this chapter the time series of all experiments are presented, showing the mass concentration evolution during each experiment. Main emphasis is put on the mass spectral differences for the different precursors and the  $\text{NO}_3^+$  ratio detected and comparing them to previous studies.

## 5.1 Time series

In this section the time series and mass concentrations for organics, nitrates and the nitrate to organic ratios ( $\text{NO}_3\text{:Org}$ ) of all four precursor experiments are presented in Figure 5.1. The green line represents the organics, the blue one is nitrates and the orange line is the  $\text{NO}_3\text{:Org}$  ratio. The  $\text{NO}_3\text{:Org}$  ratio for all experiments are presented in Figure 5.2. During the measurements, the pON particle concentration was stepwise increased or decreased to cover a wider range of mass concentrations and can be seen as stable plateaus in the organic and nitrate mass concentrations. Particle concentrations for the different steps are shown in Table 5.1. All of the data were measured with 1 minute time resolution.

**Table 5.1:** Measurement steps: showing the selected particle size and number concentration for different steps for all pON precursor VOCs.

	Acenaphthylene	Guaiacol	Limonene	$\beta$ -pinene
$d_a$ (nm)	300	200	300	400
Step 1 ( $\text{cm}^{-3}$ )	800	3200	480	400
Step 2 ( $\text{cm}^{-3}$ )	1500	7300	750	250
Step 3 ( $\text{cm}^{-3}$ )	2800	11000	1100	180
Step 4 ( $\text{cm}^{-3}$ )	3500	18000	1600	-
Step 5 ( $\text{cm}^{-3}$ )	-	26000	-	-

### 5.1.1 Acenaphthylene

The time series of acenaphthylene + NO<sub>3</sub> measurement from 11 Dec 2018 is presented in Figure 5.1a. Particle size was selected to 300 nm. The particle concentration was increased stepwise ranging from  $8 \cdot 10^2$  to  $3.5 \cdot 10^3$  cm<sup>-3</sup>. The organic and nitrate mass concentrations was measured to range from 1.7 to 9.9 and 1.0 to 5.4 µg m<sup>-3</sup>, respectively. The NO<sub>3</sub>:Org ratio was stable during the whole experiment:  $0.55 \pm 0.010$  (one standard deviation).

### 5.1.2 Guaiacol

The time series of guaiacol + NO<sub>3</sub> measurements from 7 Dec 2018 is presented in Figure 5.1c. Particle size was selected to 200 nm. The particle concentration was increased stepwise ranging from  $3.2 \cdot 10^3$  to  $2.6 \cdot 10^5$  cm<sup>-3</sup>. The organic and nitrate mass concentrations was measured to range from 4.1 to 47 and 0.59 to 5.9 µg m<sup>-3</sup>, respectively. The NO<sub>3</sub>:Org ratio was  $0.14 \pm 0.050$  (one standard deviation)

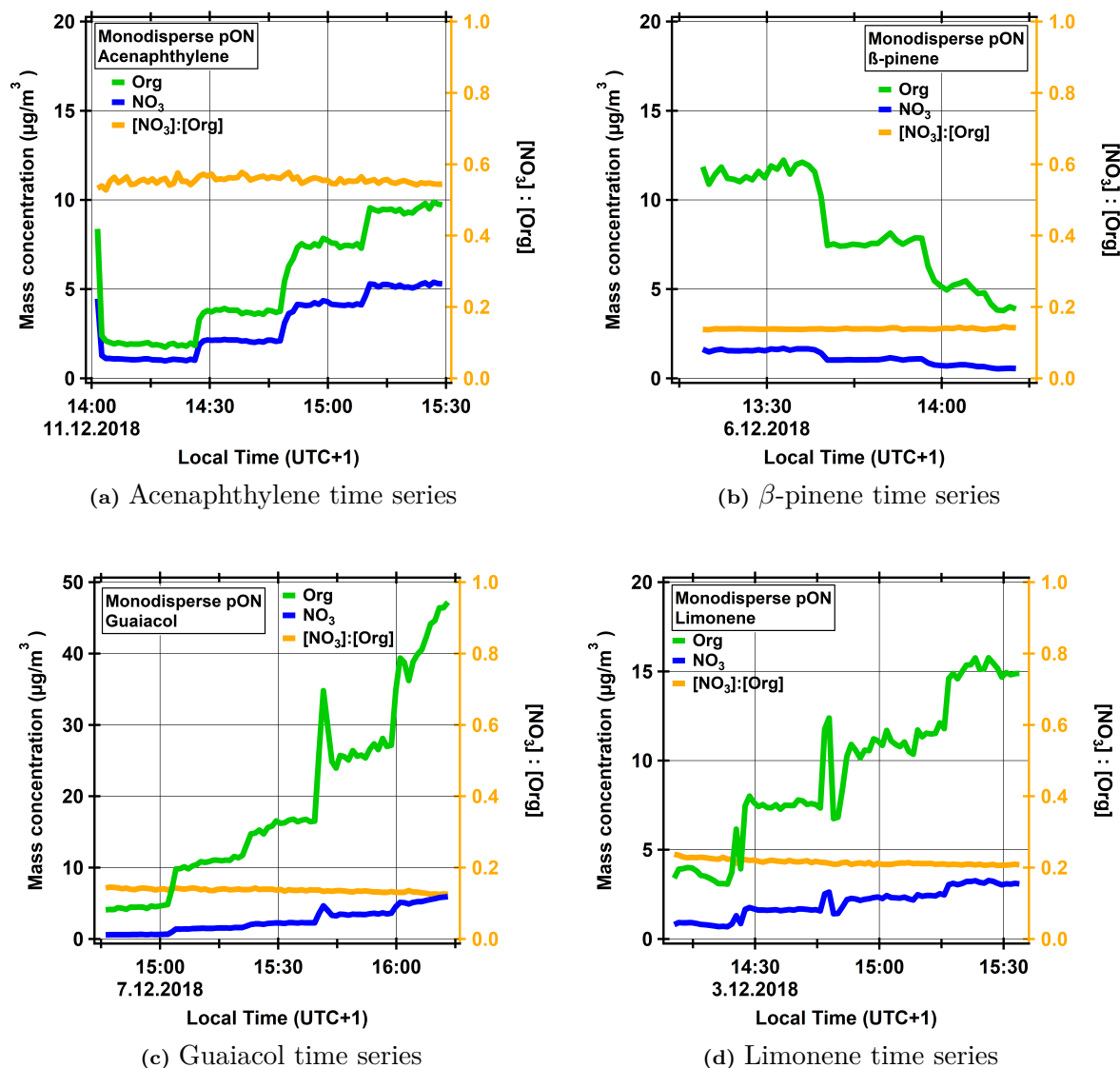
### 5.1.3 Limonene

The time series of limonene + NO<sub>3</sub> measurements from 3 Dec 2018 is presented in Figure 5.1d. Particle size was selected to 300 nm. The particle concentration was increased stepwise ranging from  $4.8 \cdot 10^2$  to  $1.6 \cdot 10^3$  cm<sup>-3</sup>. The organic and nitrate mass concentrations was measured to range from 3.1 to 16 and 0.69 to 3.3 µg m<sup>-3</sup>, respectively. The NO<sub>3</sub>:Org ratio was  $0.21 \pm 0.007$  (one standard deviation).

### 5.1.4 β-pinene

The time series of β-pinene + NO<sub>3</sub> measurements from 6 Dec 2018 is presented in Figure 5.1b. Particle size was selected to 400 nm. The particle concentration was decreased stepwise ranging from  $4.0 \cdot 10^2$  to  $1.8 \cdot 10^2$  cm<sup>-3</sup>. The organic and nitrate mass concentrations was measured to range from 12 to 3.8 and 1.7 to 0.54 µg m<sup>-3</sup>, respectively. The NO<sub>3</sub>:Org ratio was  $0.14 \pm 0.002$  (one standard deviation).

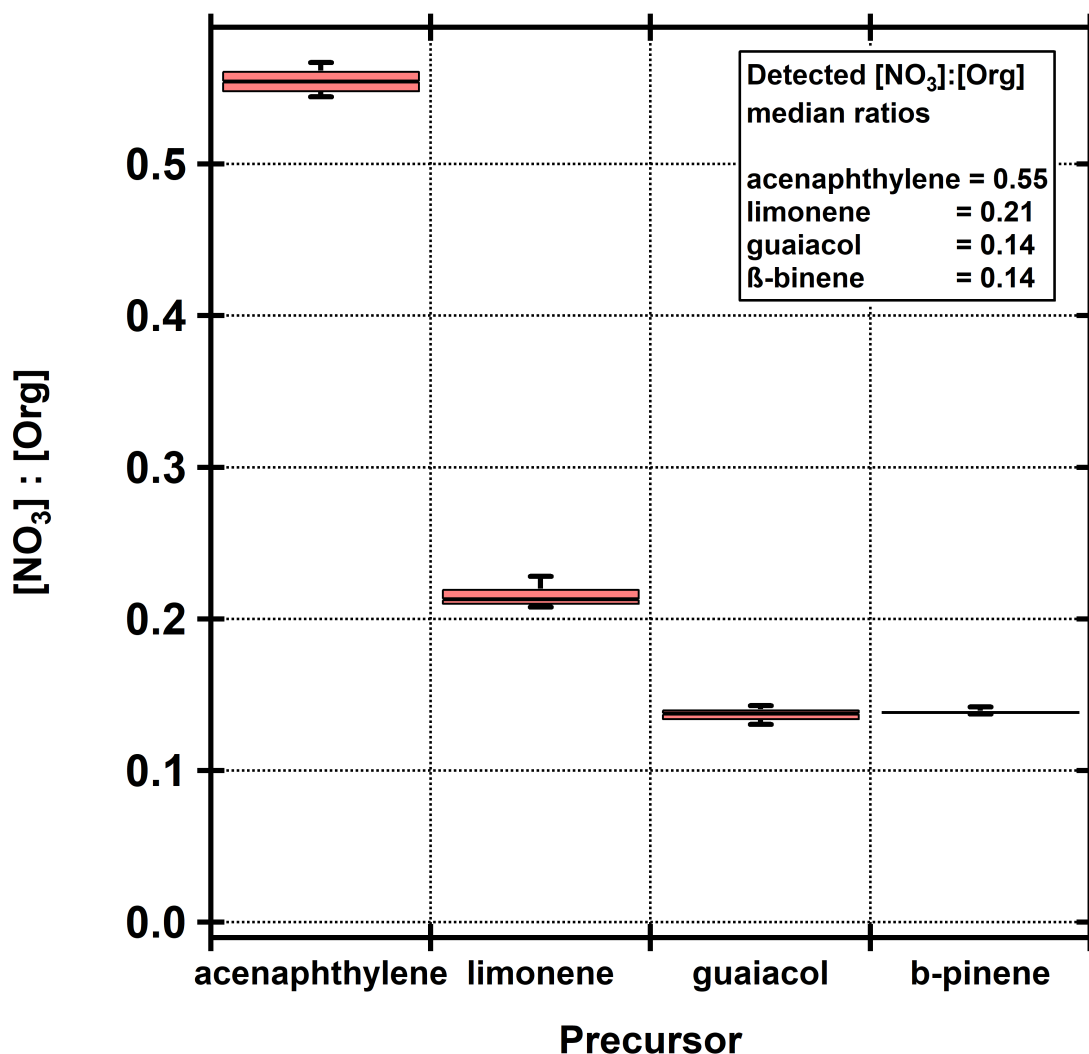




**Figure 5.1:** Time series of all four precursor experiments, showing the mass concentration of organics and nitrates and the organic to nitrate ratio. a) is for acenaphthylene, b) for  $\beta$ -pinene, c) for guaiacol and d) for limonene.

## 5.2 Mass spectral differences

The individual mass spectrum of all four precursor experiments are presented in Figures 5.3 to 5.6 and in Figure 5.7 all four mass spectrum are in the same figure to ease the comparison. The fraction of every organic family, CH, CHO1, CHOgt1, CHN, CHO1N, CHOgt1N (were gt1 stand for *greater than 1*, meaning that in those fragments there are more than one oxygen atom), is shown in Figure 5.8. In the organic families, the number of carbon, hydrogen and nitrogen atoms are not specified whereas the number of oxygen atoms is specified as one or more than one. The nitrate to organic ratio for all four experiments are shown in Figure 5.2. All of the



**Figure 5.2:** Median nitrate to organic ratio. The boxes represent the 25<sup>th</sup> and 75<sup>th</sup> percentile, whiskers are the 10<sup>th</sup> and 90<sup>th</sup> percentile.

nitrogen-containing organic fragments ( $C_xH_yO_zN_p$  fragments, where  $x, y, z, p > 0$ ) detected are listed in Tables 5.2 to 5.4, where the fragments coloured in red are unique fragments that are detected only for one precursor and could possibly be used as tracers. In Appendix A, there are a few screenshots from the data analysis part to see how well the L-ToF-AMS is able to separate fragments with small mass-difference. From there it is also seen that most of the  $C_xH_yO_zN_p$  fragments are very small (often only a few percent of the total signal at its unit mass) and close to other fragments, and even higher resolution is needed to separate these.

The acenaphthylene +  $NO_3$  mass spectrum (Figure 5.3) is dominated by simple hydrocarbon (CH) fragments, although the  $NO^+$  fragment is the largest single peak with its over 20% relative contribution to the total detected signal. The CH fragments

accounts for nearly 70% of the total organic signal, while the contribution the nitrogen containing families, CHN, CHNO1 and CHOgt1N, the contributions are small: 0.3%, 0.7% and 0.7%, respectively. Unlike the other investigated precursors, acenaphthylene has many of its organic fragments at high  $m/z$  values: several CH and some CHO1 fragments are at  $m/z > 110$ . Nitrogen-containing organic fragments that are a major peak at its unit mass are found at  $m/z = 57, 58, 69$  and  $70$ . Acenaphthylene has the highest nitrate to organic ratio, 0.55, indicating that a large fraction of its pON is fragmented to  $\text{NO}^+$  and  $\text{NO}_2^+$ .

In the  $\beta$ -pinene +  $\text{NO}_3$  organic mass spectrum (Figure 5.4), CH and CHO1 fragments (69% and 28%) accounts for over 96% of all signal, leaving the contribution of CHOgt1, CHN, CHO1N and CHOgt1 fragments to 2.6%, 0.3%, 0.08% and 0.3%. Similar to acenaphthylene,  $\text{NO}^+$  is the largest single fragment. However, its relative contribution is only 8% of the total signal. Together with guaiacol,  $\beta$ -pinene has the lowest nitrate to organic ratio, 0.14 and no major  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$  fragments were detected.

The limonene +  $\text{NO}_3$  organic mass spectrum (Figure 5.5) is dominated by CH and CHO1 fragments, they contribute 50% and 46% of all organic signal. The high CHO1 fraction is due to the large peak at  $m/z$  43 and the  $\text{C}_2\text{H}_3\text{O}^+$  fragment, it is the largest single fragment detected and stands alone for almost 20% of the total signal in the mass spectra. CHOgt1 accounts for 3% and all the nitrogen-containing families contribute 0.8% each. Limonene produced least  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$  fragments, only 14 fragments were detected. In addition to this, all of its  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$  fragments were detected at such unit masses where  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$  fragments for the other precursors were also detected, so limonene as pON precursor can not be identified by these fragments. The nitrate to organic ratio was 0.21.

The guaiacol +  $\text{NO}_3$  organic mass spectrum (Figure 5.6) is the only where the CH is not the largest organic family, here the largest family is CHO1 with its 38% contribution, while CH stands for 27% and CHOgt1 26% of the organics. The high CHO1 fraction is mainly due to the fragments at  $m/z$  28 ( $\text{CO}^+$ ) and 29 ( $\text{CHO}^+$ ), each accounts for 7 to 8% of the total signal in the guaiacol mass spectra. Guaiacol has by far the highest amount of nitrogen containing fragments: CHN, CHO1N and CHOgt1N accounts for 4%, 1% and 0.6% of the organics. From this experiment, the largest amount (over 40) of nitrogen-containing organic fragments were detected, but only at  $m/z = 67, 68$  and  $80$  it was the major fragment at that unit mass. The rest of the  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$  fragments did not contribute significantly to the signal at its unit mass. The nitrate to organic ratio was 0.14. The CHOgt1N fragments in the organic mass

spectra (Figure 5.6) looks unusual, especially as it contains several fragments at  $m/z > 130$ . Although there are many of these fragments, their contribution to the total organic mass spectrum is, as earlier mentioned, only 0.6%, meaning that these fragments are small. One explanation to the vast amount of these CHOgt1N fragments might be related to errors in the peak shape determination or the  $m/z$  calibration during the data analysis. This is possible because during the HR-analysis one is required to choose every fragment that are being fitted to the collected raw data. If the raw data signal is noisy or several fragments are close each other, such errors may happen. This opens up the possibility for errors and leads to choosing fragments that actually are not there because the raw data is not properly processed. Further work will explore the validity of these results in more detail.

### 5.3 $\text{NO}_x^+$ ratios

In this section, the detected  $\text{NO}^+$  to  $\text{NO}_2^+$  ratio ( $\text{NO}_x^+$  ratio) for all four precursors are presented in Figure 5.9. For all four precursors, a straight line was fitted to the  $\text{NO}^+$  and  $\text{NO}_2^+$  data to get the  $\text{NO}_x^+$  ratio as the slope of the fit. The linear fit is very good (Pearson correlation coefficient,  $r$ , is over 0.99 for all four cases) for all four precursors, as shown in Figure 5.11.

The median  $\text{NO}_x^+$  ratio was quite similar for all four precursors, the highest  $\text{NO}_x^+$  ratio was measured for acenaphthylene (6.70), followed by guaiacol (6.60),  $\beta$ -pinene (6.23) and limonene (5.96). When comparing our measured  $\text{NO}_x^+$  ratios with previous studies with same precursors and  $\text{NO}_3$  oxidation (see Figure 5.10), large variation is seen especially for  $\beta$ -pinene. All other studies were carried out with a HR-ToF-AMS, also taking into account the  $\text{CH}_2\text{O}^+$  fragment at  $m/z$  30 when calculating the  $\text{NO}^+$  signal. One reason to the variation might be instrumental bias, discussed already in [Boyd et al., 2015], and as no previous study has been done with the L-ToF-AMS, a new instrumental bias might occur. For acenaphthylene there were no previous studies, but the purity of only 75% (rest acenaphthene) must be remembered when future studies are compared to these results.

**Table 5.2:** Detected  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$  fragments between  $m/z$  43 and 89. Fragments coloured in red are unique fragments that are detected only for one precursor. All of the fragments are positive ions.

$m/z$	Acenaphthylene	Guaiacol	Limonene	$\beta$ -pinene
43	CHNO	CHNO		
44				
45			$\text{CH}_3\text{NO}$	$\text{CH}_3\text{NO}$
46				
47			$\text{CH}_5\text{NO}$	$\text{CH}_5\text{NO}$
48				
54		$\text{C}_2\text{HNO}$		
55	$\text{C}_2\text{HNO}$			
56	$\text{C}_2\text{H}_2\text{NO}$	$\text{C}_2\text{H}_2\text{NO}$		
57	$\text{C}_4\text{H}_6\text{NO}_2$	$\text{C}_2\text{H}_3\text{NO}$		$\text{C}_2\text{H}_3\text{NO}$
58	$\text{C}_2\text{H}_4\text{NO}$	$\text{C}_2\text{H}_4\text{NO}$		$\text{C}_2\text{H}_4\text{NO}$
59				$\text{C}_2\text{H}_5\text{NO}$
60		$\text{CH}_2\text{NO}_2$	$\text{C}_2\text{H}_6\text{NO}$ $\text{CH}_2\text{NO}_2$	$\text{C}_2\text{H}_6\text{NO}$ $\text{CH}_2\text{NO}_2$
61		$\text{CH}_3\text{NO}_2$		
62				$\text{CH}_4\text{NO}_2$
63				$\text{CH}_5\text{NO}_2$
64				
67	$\text{C}_3\text{HNO}$	$\text{C}_3\text{HNO}$		
68	$\text{C}_3\text{H}_2\text{NO}$	$\text{C}_3\text{H}_2\text{NO}$		
69	$\text{C}_3\text{H}_3\text{NO}$			
70	$\text{C}_3\text{H}_4\text{NO}$	$\text{C}_3\text{H}_4\text{NO}$		
71				$\text{C}_3\text{H}_5\text{NO}$
72		$\text{C}_2\text{H}_2\text{NO}_2$		$\text{C}_3\text{H}_6\text{NO}$
73				$\text{C}_2\text{H}_5\text{N}_2\text{O}$
74				
75				$\text{C}_2\text{H}_5\text{NO}_2$
76			$\text{CH}_2\text{NO}_3$	$\text{CH}_2\text{NO}_3$
77				
78		$\text{C}_4\text{NO}$		
79				
80		$\text{C}_4\text{H}_2\text{NO}$		
81				
82				
86		$\text{C}_3\text{H}_4\text{NO}_2$		$\text{C}_3\text{H}_4\text{NO}_2$
87	$\text{C}_4\text{H}_9\text{NO}$	$\text{C}_3\text{H}_5\text{NO}_2$		
88			$\text{C}_3\text{H}_6\text{NO}_2$	$\text{C}_3\text{H}_6\text{NO}_2$
89				

**Table 5.3:** Detected  $C_xH_yO_zN_p$  fragments between  $m/z$  90 and 139. Fragments coloured in red are unique fragments that are detected only for one precursor. All of the fragments are positive ions.

$m/z$	Acenaphthylene	Guaiacol	Limonene	$\beta$ -pinene
90		$C_2H_4NO_3$		
91				
92				
98		$C_4H_4NO_2$		
100	$C_5H_{10}NO$			
101			$C_4H_7NO_2$	$C_4H_7NO_2$
102		$C_3H_4NO_3$		
103				
104		$C_3H_6NO_3$		
105				
106				
109		$C_6H_7NO$		
110				
111				
112		$C_5H_6NO_2$		
113				
114				
117	$C_5H_{11}NO_2$			
118				
120	$C_6H_2NO_2$			
121	$C_5HN_2O_2$			
122				
128		$C_4H_2NO_4$		$C_6H_{10}NO_2$
129				
132		$C_4H_6NO_4$		$C_4H_6NO_4$
133	$C_4H_7NO_4$	$C_3HN_2O_5$		
134	$C_7H_6N_2O$			
135				
136		$C_6H_2NO_3$		
137				
138		$C_6H_4NO_3$		
139		$C_6H_5NO_3$		

**Table 5.4:** Detected  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$  fragments between  $m/z$  140 and 160. Fragments coloured in red are unique fragments that are detected only for one precursor. All of the fragments are positive ions.

$m/z$	Acenaphthylene	Guaiacol	Limonene	$\beta$ -pinene
140		$\text{C}_5\text{H}_2\text{NO}_4$ $\text{C}_6\text{H}_6\text{NO}_3$		
141	$\text{C}_5\text{H}_5\text{N}_2\text{O}_3$ $\text{C}_6\text{H}_7\text{NO}_3$	$\text{C}_5\text{H}_5\text{N}_2\text{O}_3$ $\text{C}_6\text{H}_7\text{NO}_3$ $\text{C}_7\text{H}_{11}\text{NO}_2$	$\text{C}_7\text{H}_{11}\text{NO}_2$	
142	$\text{C}_6\text{H}_8\text{NO}_3$	$\text{C}_6\text{H}_8\text{NO}_3$ $\text{C}_5\text{H}_4\text{NO}_4$	$\text{C}_6\text{H}_8\text{NO}_3$ $\text{C}_7\text{H}_{12}\text{NO}_2$	$\text{C}_6\text{H}_8\text{NO}_3$
143		$\text{C}_5\text{H}_5\text{NO}_4$	$\text{C}_6\text{H}_9\text{NO}_3$ $\text{C}_7\text{H}_{13}\text{NO}_2$	
144		$\text{C}_5\text{H}_6\text{NO}_4$		$\text{C}_5\text{H}_6\text{NO}_4$ $\text{C}_7\text{H}_{14}\text{NO}_2$
145	$\text{C}_6\text{H}_{11}\text{NO}_3$	$\text{C}_5\text{H}_7\text{NO}_4$ $\text{C}_6\text{H}_{11}\text{NO}_3$		$\text{C}_7\text{H}_{15}\text{NO}_2$
146				
147	$\text{C}_5\text{H}_9\text{NO}_4$			
148	$\text{C}_7\text{H}_2\text{NO}_3$ $\text{C}_5\text{H}_{10}\text{NO}_4$	$\text{C}_7\text{H}_2\text{NO}_3$ $\text{C}_5\text{H}_{10}\text{NO}_4$		$\text{C}_5\text{H}_{10}\text{NO}_4$
149				
150				
151		$\text{C}_7\text{H}_5\text{NO}_3$		
152		$\text{C}_6\text{H}_2\text{NO}_4$ $\text{C}_7\text{H}_6\text{NO}_3$		
153		$\text{C}_6\text{H}_3\text{NO}_4$ $\text{C}_7\text{H}_7\text{NO}_3$	$\text{C}_9\text{H}_{15}\text{NO}$	$\text{C}_9\text{H}_{15}\text{NO}$
154	$\text{C}_7\text{H}_8\text{NO}_3$	$\text{C}_7\text{H}_8\text{NO}_3$ $\text{C}_6\text{H}_4\text{NO}_4$		
155	$\text{C}_7\text{H}_9\text{NO}_3$	$\text{C}_7\text{H}_9\text{NO}_3$ $\text{C}_6\text{H}_5\text{NO}_4$		$\text{C}_7\text{H}_9\text{NO}_3$
156	$\text{C}_7\text{H}_{10}\text{NO}_3$	$\text{C}_7\text{H}_{10}\text{NO}_3$ $\text{C}_6\text{H}_6\text{NO}_4$	$\text{C}_7\text{H}_{10}\text{NO}_3$	
157		$\text{C}_6\text{H}_7\text{NO}_4$	$\text{C}_7\text{H}_{11}\text{NO}_3$	$\text{C}_7\text{H}_{11}\text{NO}_3$
158		$\text{C}_6\text{H}_8\text{NO}_4$	$\text{C}_6\text{H}_8\text{NO}_4$	$\text{C}_6\text{H}_8\text{NO}_4$
159		$\text{C}_6\text{H}_9\text{NO}_4$		$\text{C}_6\text{H}_9\text{NO}_4$

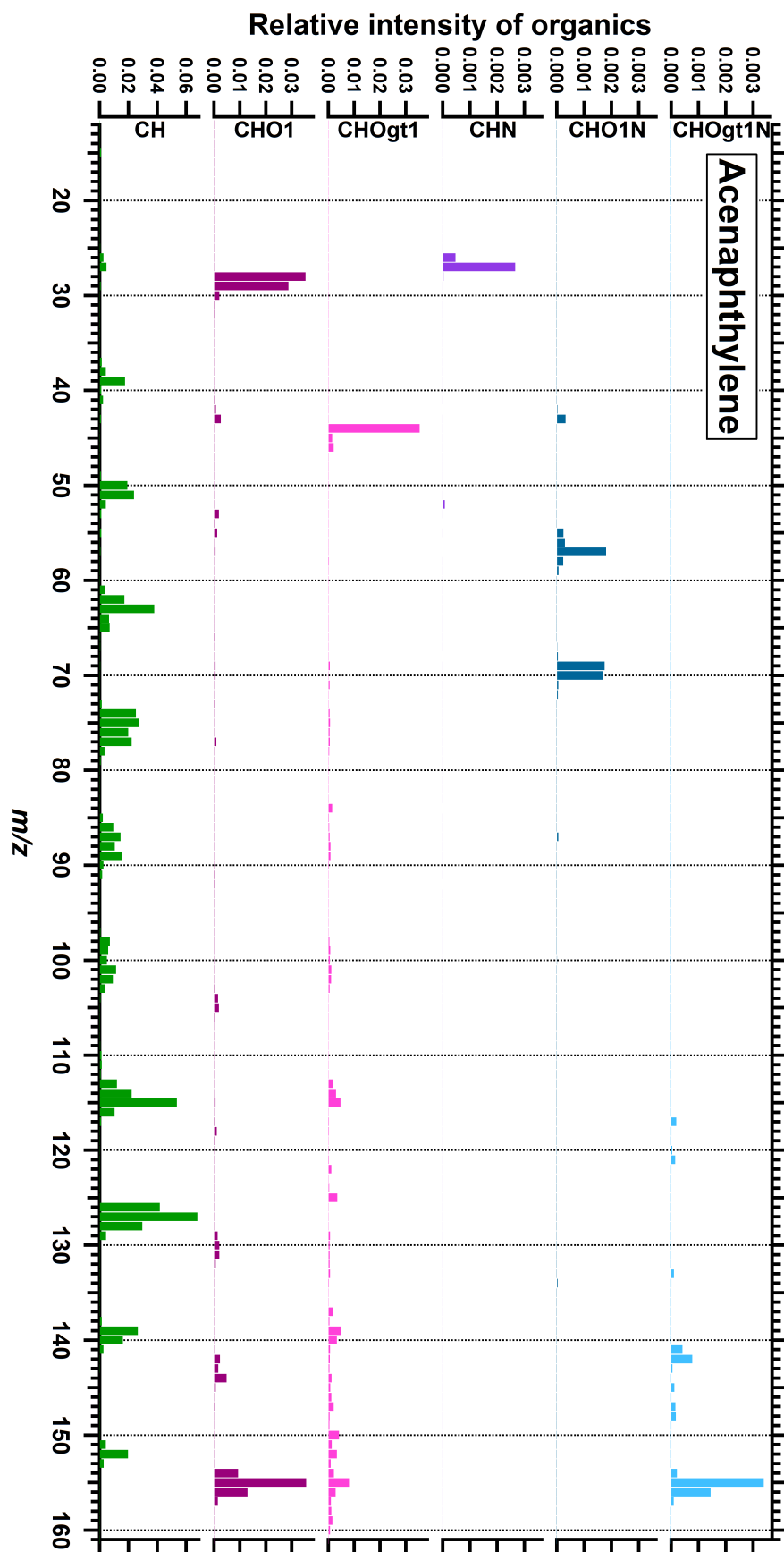


Figure 5.3: Mass spectrum of organic fragments from the acenaphthylene +  $\text{NO}_3$  experiment. Mass spectrum separated by organic family.



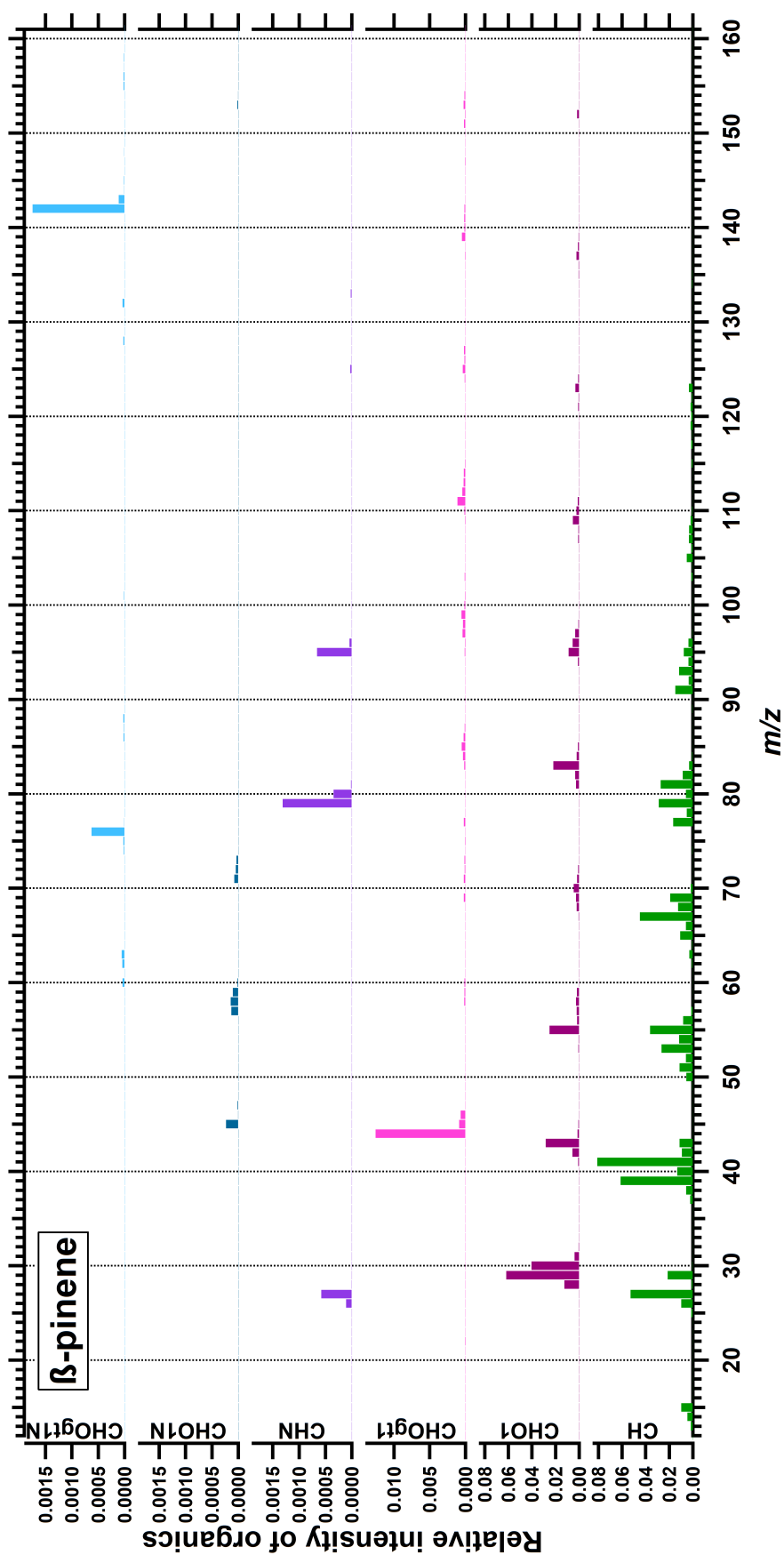


Figure 5.4: Mass spectrum of organic fragments from the  $\beta$ -pinene +  $\text{NO}_3$  experiment. Mass spectrum separated by organic family.

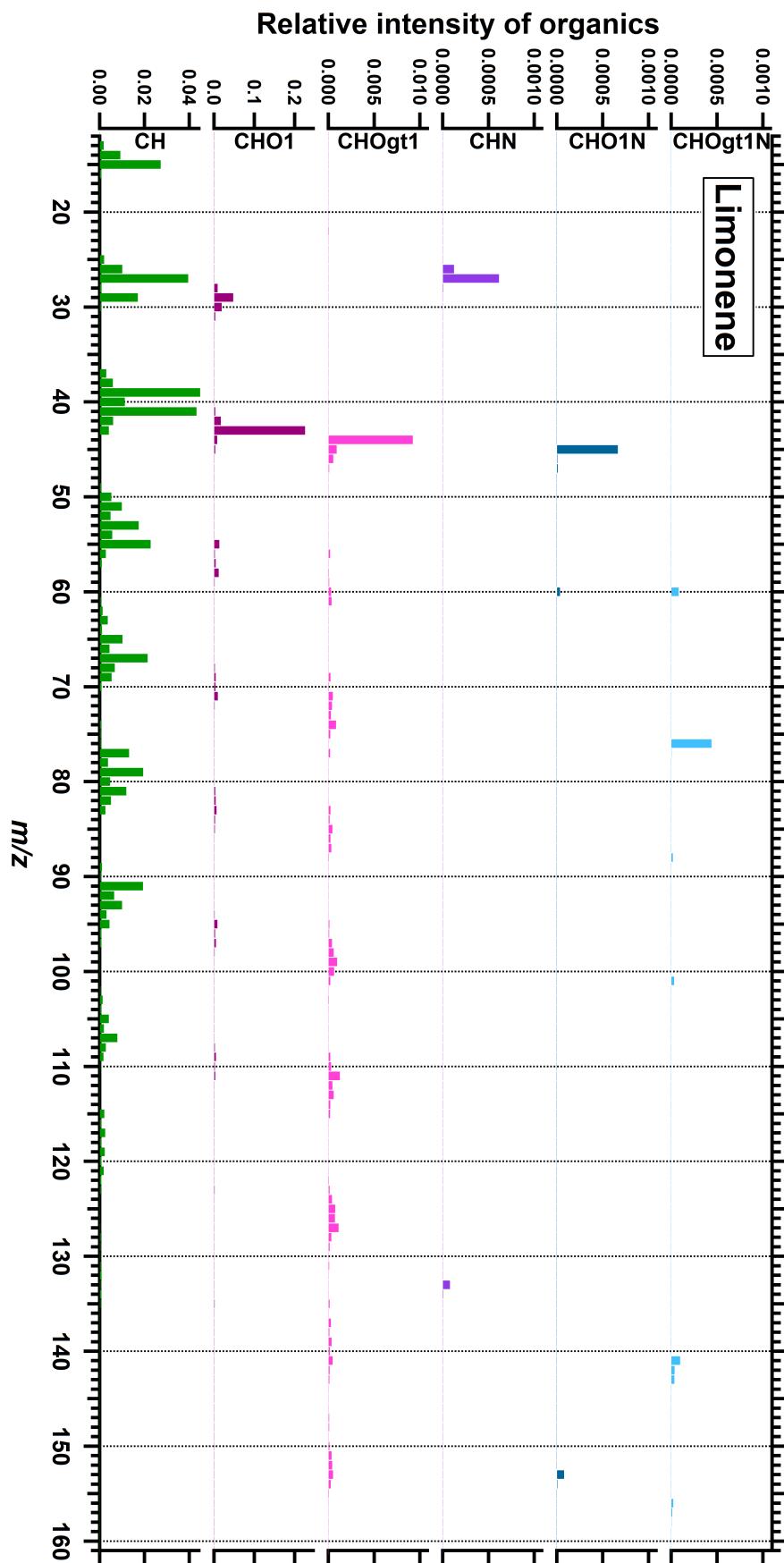


Figure 5.5: Mass spectrum of organic fragments from the limonene +  $\text{NO}_3$  experiment. Mass spectrum separated by organic family.

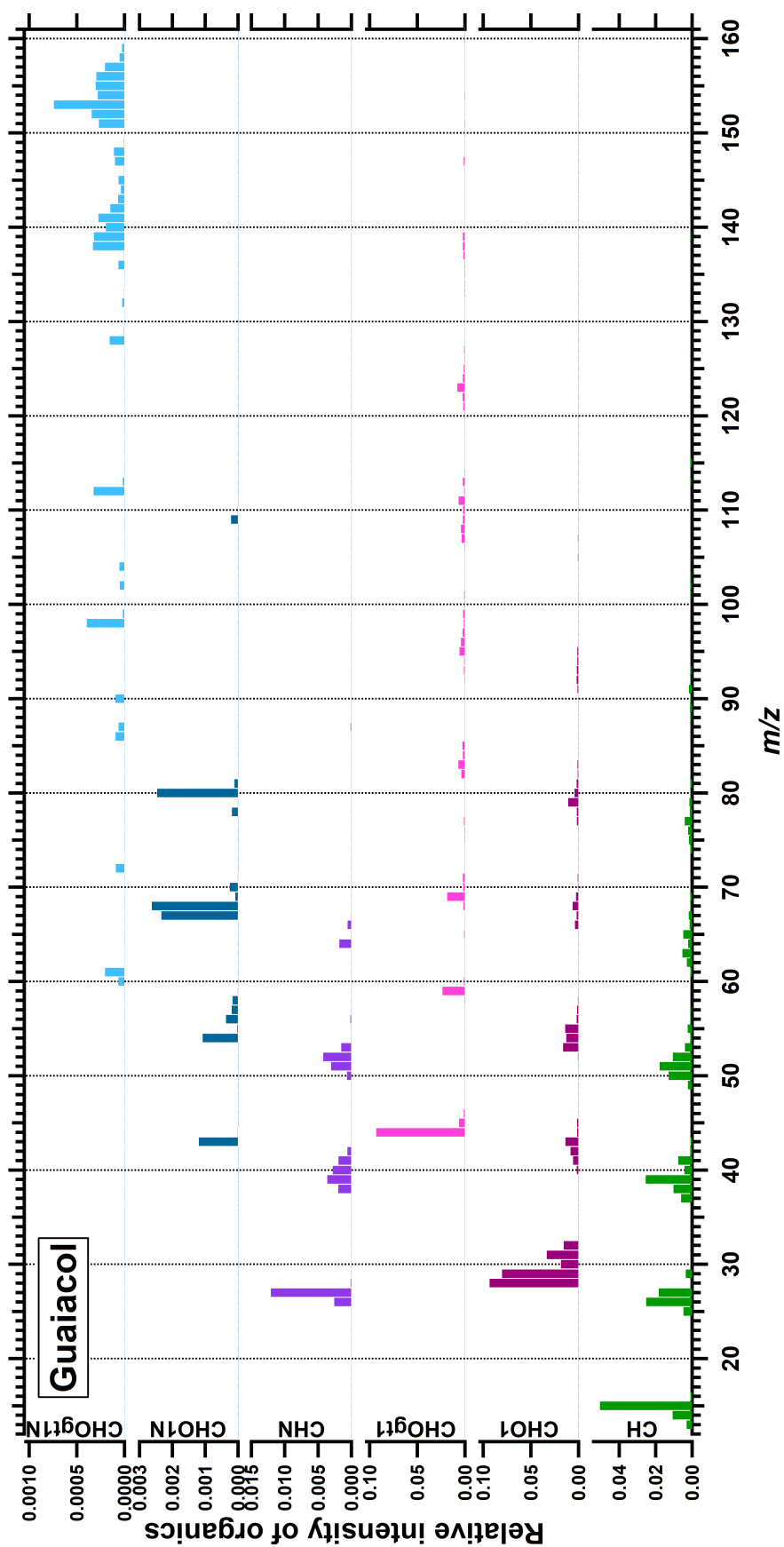


Figure 5.6: Mass spectrum of organic fragments from the guaiaicol +  $\text{NO}_3$  experiment. Mass spectrum separated by organic family.

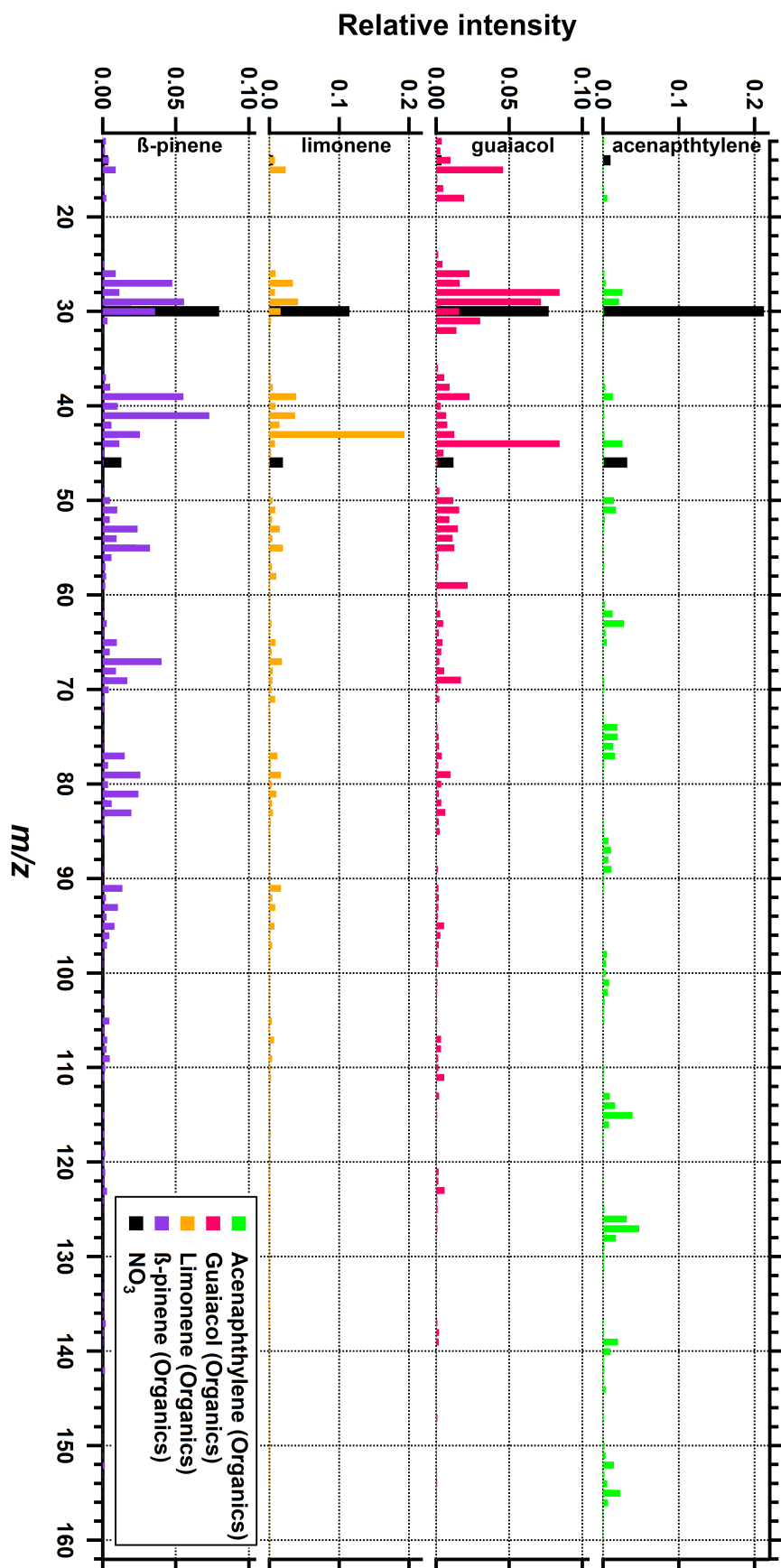


Figure 5.7: Mass spectrum of all four precursor experiments.

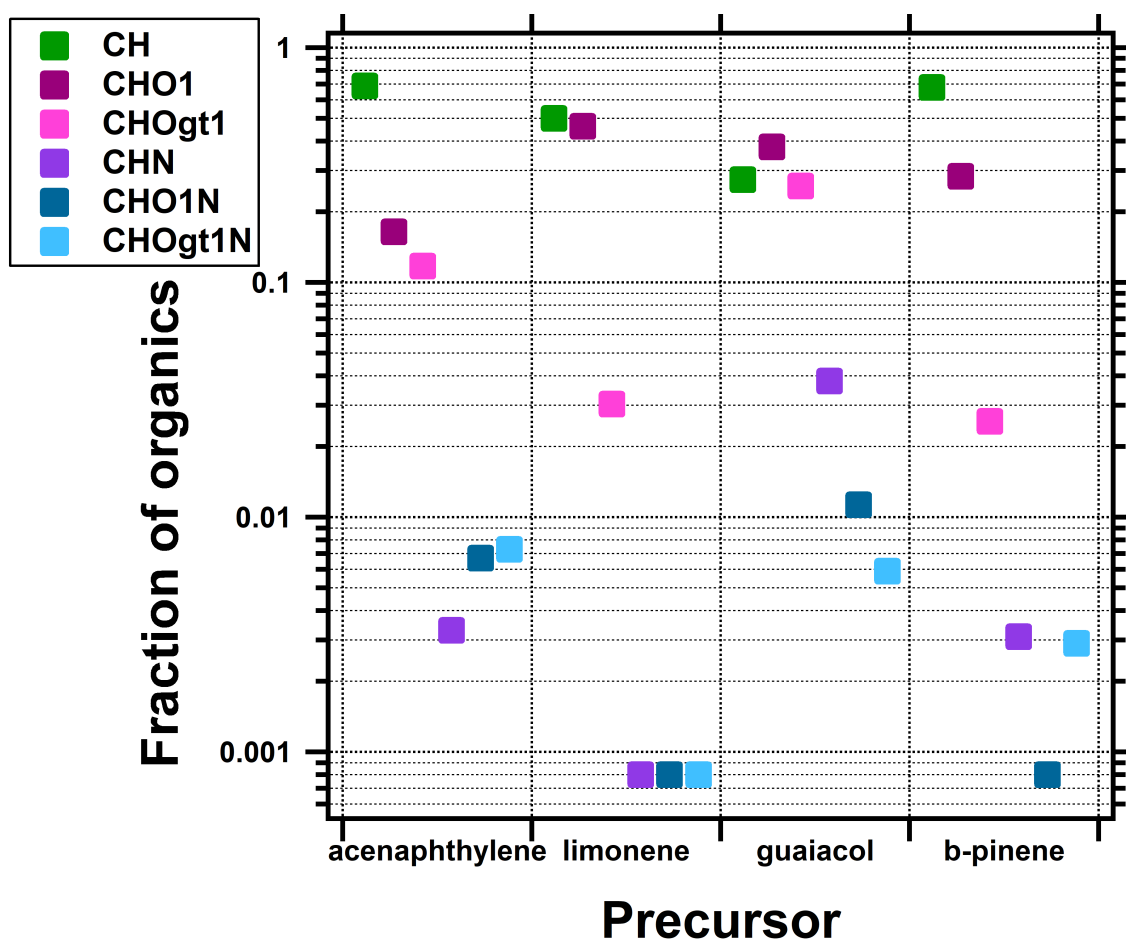
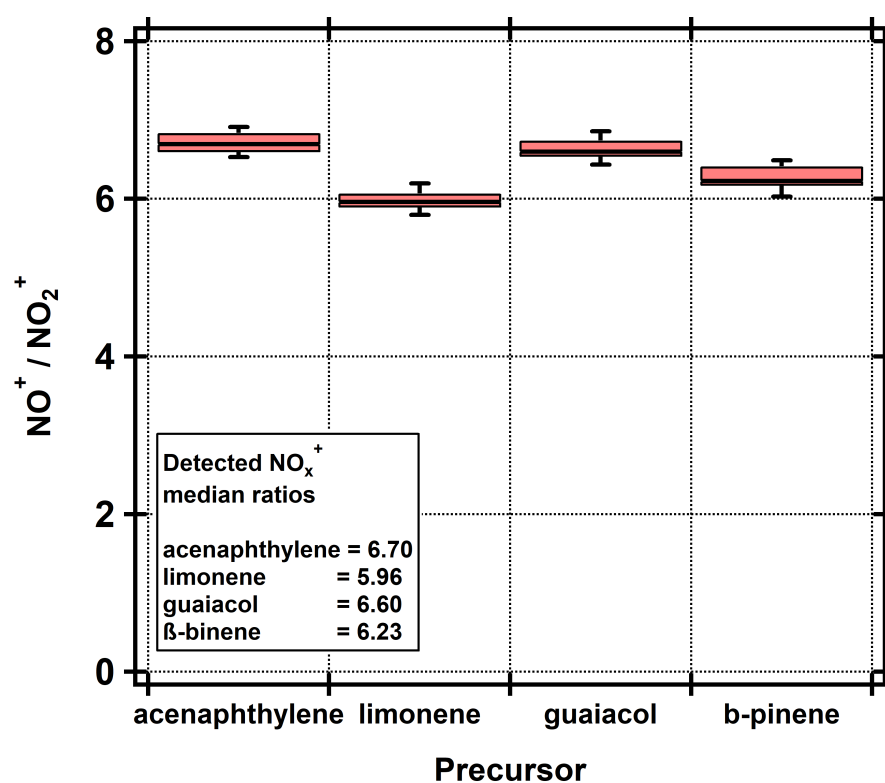
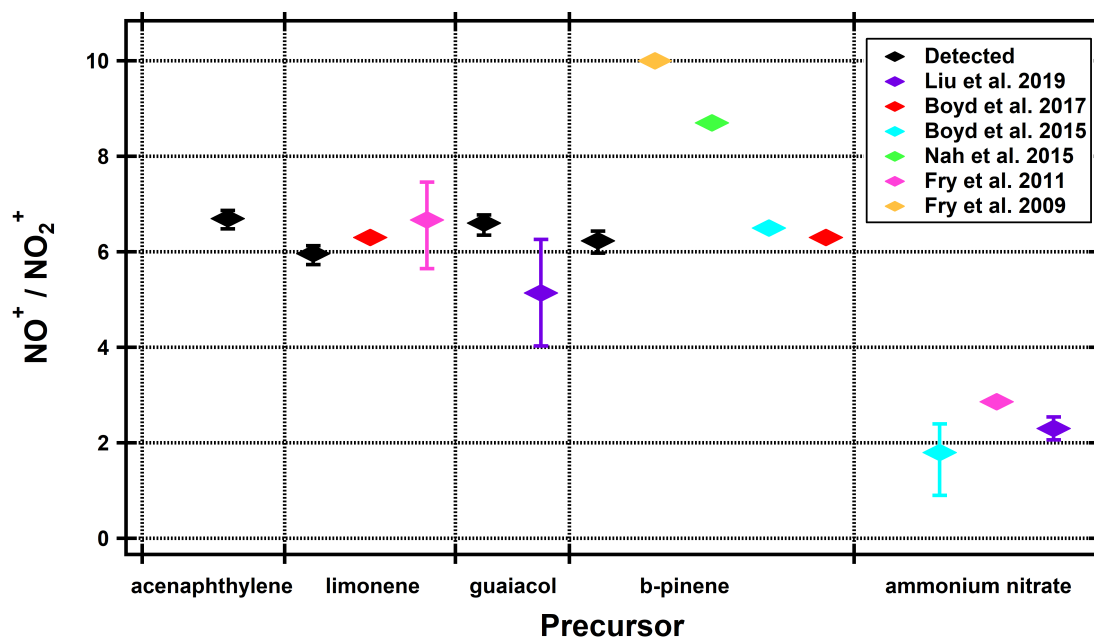


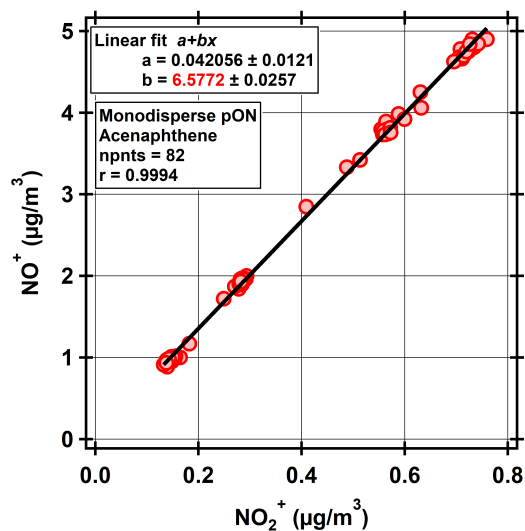
Figure 5.8: The organic families fraction of the total organic signal for each precursor.



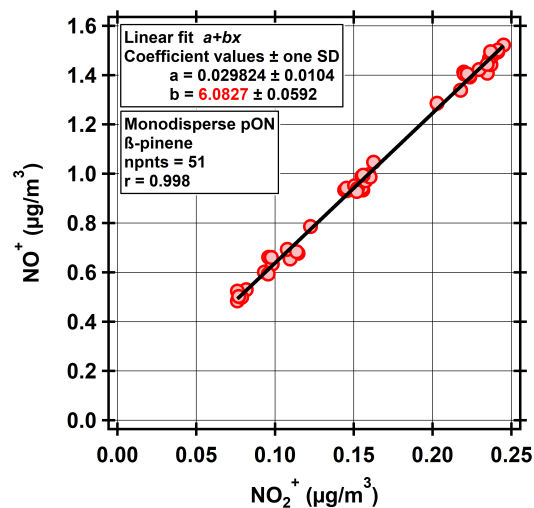
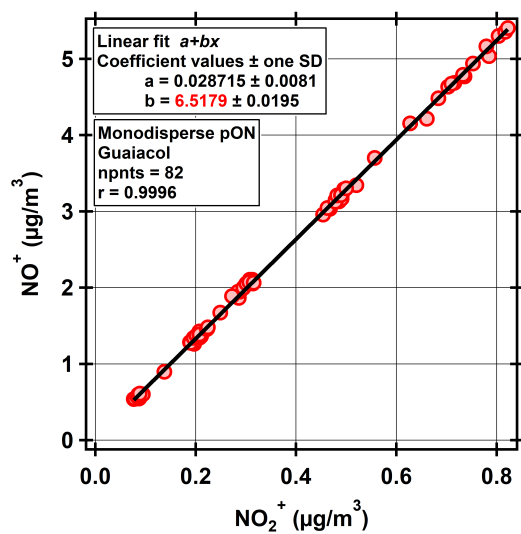
**Figure 5.9:** Detected median values for the  $\text{NO}_x^+$  ratio for all 4 precursors. The boxes represent the 25<sup>th</sup> and 75<sup>th</sup> percentile, whiskers are the 10<sup>th</sup> and 90<sup>th</sup> percentile.



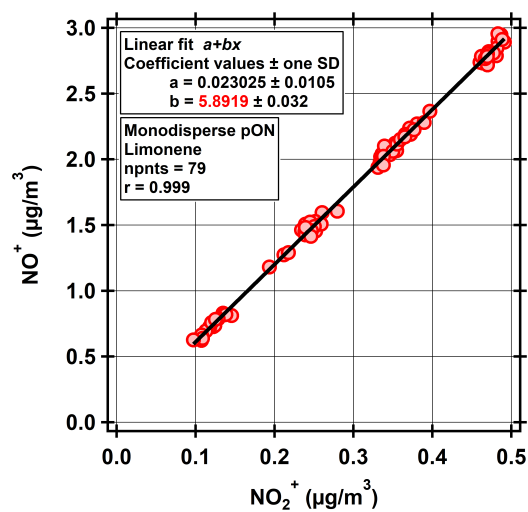
**Figure 5.10:** Detected and literature values for  $\text{NO}_x^+$  ratios for all 4 precursors and ammonium nitrate. Black markers are values measured during this work, rest is previous studies.



(a) Acenaphthylene

(b)  $\beta$ -pinene

(c) Guaiacol



(d) Limonene

Figure 5.11:  $\text{NO}_x^+$  ratio for all four precursors.





## 6. Conclusions

At the ACMCC, we generated monodisperse pON in a PAM-chamber by oxidizing four different VOCs with  $\text{NO}_3$  radicals and sampled it with 8 ACSM instruments as well as a L-ToF-AMS that I operated. I investigated the fragments produced in the AMS by utilizing the high resolution of the L-ToF-AMS, and was able to separate fragments with different elemental composition and divide them into different organic families. VOCs used in the work was  $\beta$ -pinene, limonene, guaiacol and acenaphthylene.

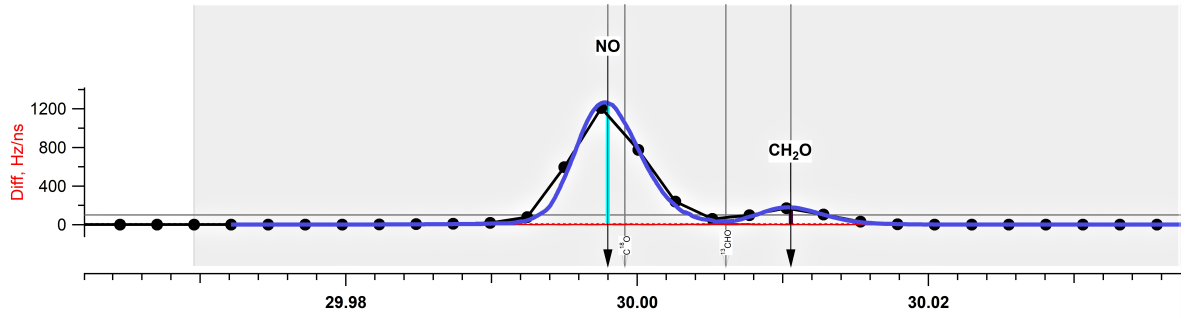
Using the  $\text{NO}_x^+$  ratio for separating and quantifying organic nitrate from inorganic nitrate has been previously suggested, and in this work we measured quite similar  $\text{NO}_x^+$  ratio for all four pON-precursors. Our measured  $\text{NO}_x^+$  ratios are in good consistent with most previous studies, although some studies differ from our results. The measured  $\text{NO}_x^+$  ratio in our experiments for all four precursors are within the range of  $6.3 \pm 0.4$ .

The main focus in this work was investigating the differences in the fragmentation patterns for the different precursors and detecting  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$ -fragments, something that has been previously challenging due to limitations of the AMSs resolution. During this work, I tentatively detected over 100 different  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$ -fragments, which can now possibly be used as marker fragments for these precursors in ambient measurements. Especially for the guaiacol and acenaphthylene experiments, I found several unique  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$ -fragments. Although the L-ToF-AMS has higher resolution than any older AMS version and is capable of separating  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$ -fragments, there are still some uncertainties, mainly linked to possible problems with the  $m/z$  calibration and the peak shape determination during the data analysis. Because the  $\text{C}_x\text{H}_y\text{O}_z\text{N}_p$ -fragments are often very small and close to some other fragment, they might appear in the data analysis even though they would not actually be present. Actual fragments might also be lost with this kind of problems. This is an aspect that always must be considered when analysing L-ToF-AMS data.

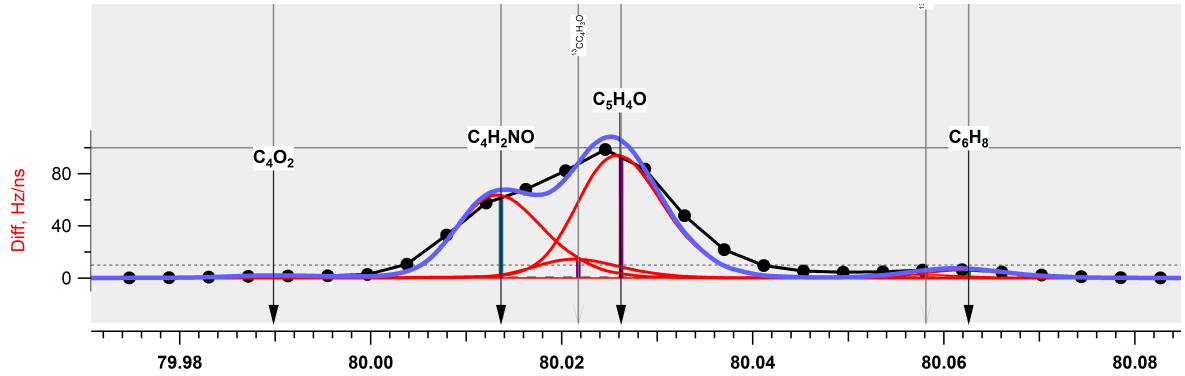
As shown in this and previous studies, the oxidation of BVOCs with  $\text{NO}_3$  radicals are a direct link between natural and anthropogenic emissions and aerosol formation. These results can also be utilized in the improvement of the fragmentation table for UMR instruments (for example the ACSM) and source apportionment of SOA formation.

Future research should focus on quantifying the aerosol yield from BVOC + NO<sub>3</sub> reactions and studying the particle phase evolution of pON. In this work, we used only particle phase instruments, but combining these results with gas-phase measurements could give us crucial new information about pON and NO<sub>x</sub> cycling in the atmosphere and reduce the gap between measurements and theory. This knowledge is essential for the quantification of pON yields.

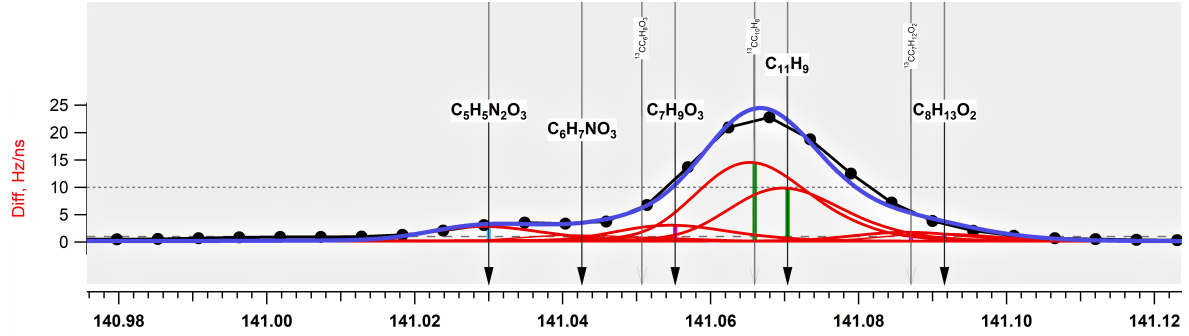
# A. Peak fitting during L-ToF-AMS data analysis



**Figure A.1:** Screenshot from peak-fitting during the data-analysis when choosing which fragments contributes to each signal. At  $m/z = 30$  from the limonene +  $\text{NO}_3$  experiment, we have the  $\text{NO}^+$  and  $\text{CH}_2\text{O}^+$  fragments. The black dotted line represents the raw data from the AMS and the blue one is the fitted sum-line for all fragments.



**Figure A.2:** Screenshot from peak fitting during the data analysis when choosing which fragments contributes to each signal. At  $m/z = 80$  from the guaiacol +  $\text{NO}_3$  experiment, we have  $\text{C}_4\text{H}_2\text{NO}^+$  and  $\text{C}_5\text{H}_4\text{O}^+$  as the main fragments. The black dotted line represents the raw data from the AMS, the red ones are fitted peaks for each fragment and the blue line is the fitted sum-line for all fragments. We can see here that the fitted blue line do not perfectly line up with the black raw data.



**Figure A.3:** Screenshot from peak fitting during the data analysis when choosing which fragments contributes to each signal. At  $m/z = 141$  from the acenaphthylene +  $\text{NO}_3$  experiment, we have several fragments that can not be separated. The black dotted line represents the raw data from the AMS, the red ones are fitted peaks for each fragment and the blue line is the fitted sum-line for all fragments. Here we can see that the individual fragments can not be separated, but summed up together they represent the detected raw data.

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